

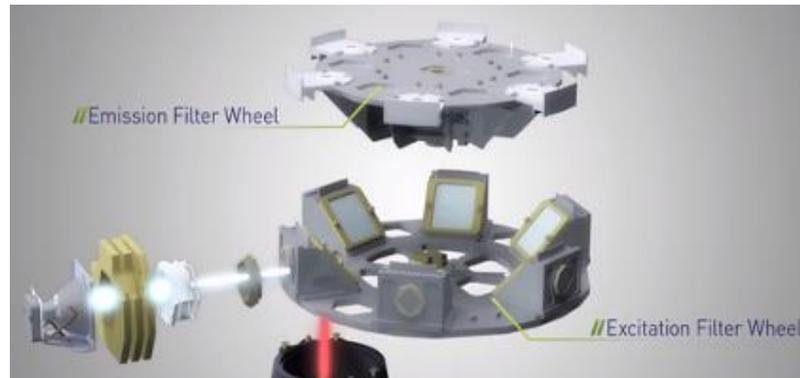


QuantStudio qPCR System Overview

Scott.Gardner@thermofisher.com
Field Applications Scientist

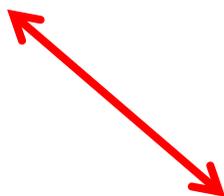
The Optics: Enhanced OptiFlex™ System

- light source
- CCD Camera captures data of all filters at every cycle



	m1(520±15)	m2(558±11)	m3(586±10)	m4(623±14)	m5(682±14)	m6(711±12)
x1(470±15)	<input checked="" type="checkbox"/>	<input type="checkbox"/>				
x2(520±10)		<input type="checkbox"/>				
x3(550±11)			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
x4(580±10)				<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
x5(640±10)					<input type="checkbox"/>	<input type="checkbox"/>
x6(662±10)						<input type="checkbox"/>

QuantStudio Connectivity

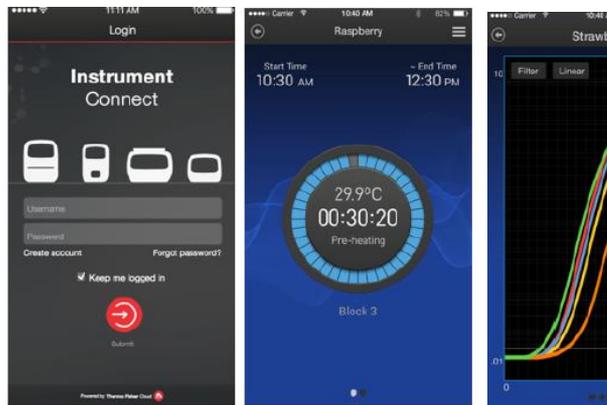


USB

Thermo Fisher Connect™



PC or Mac®
Connect with a web browser

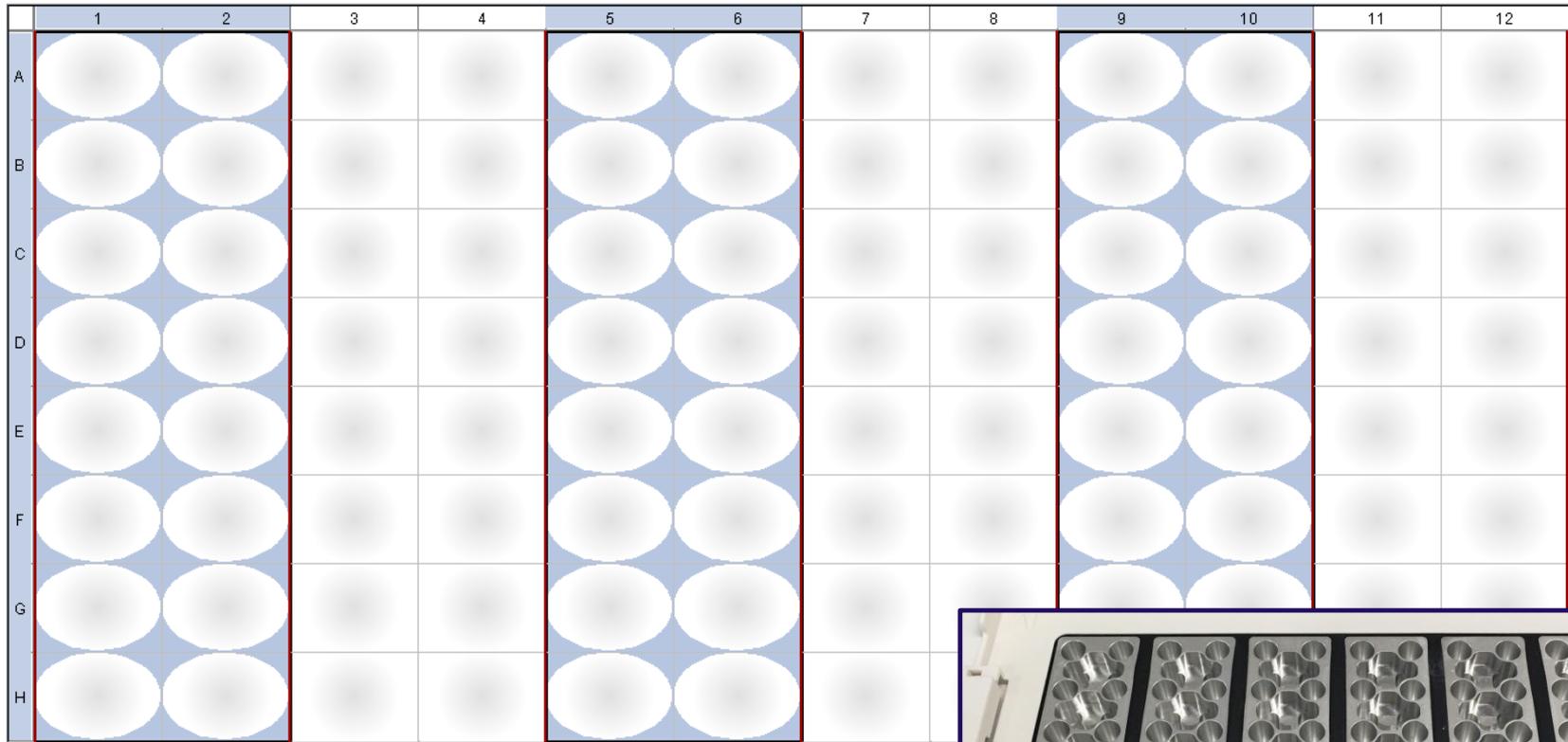


Connect to your data anytime, anywhere

ThermoFisher
SCIENTIFIC

VeriFlex™ Block technology

6 independently controlled programmable zones ($\leq 5c$ of adjacent zone)



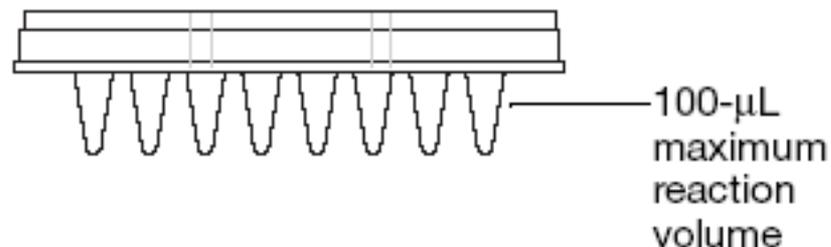
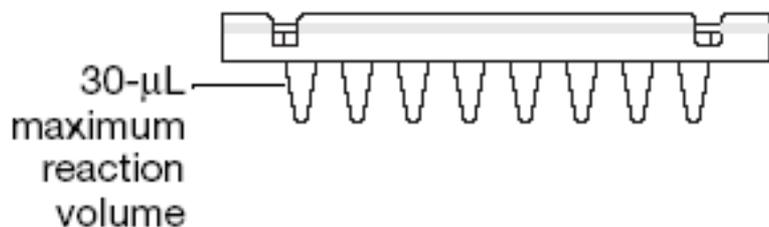
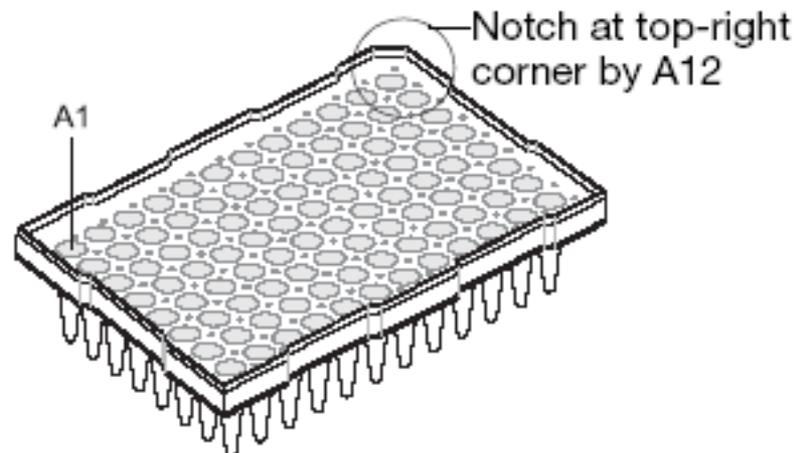
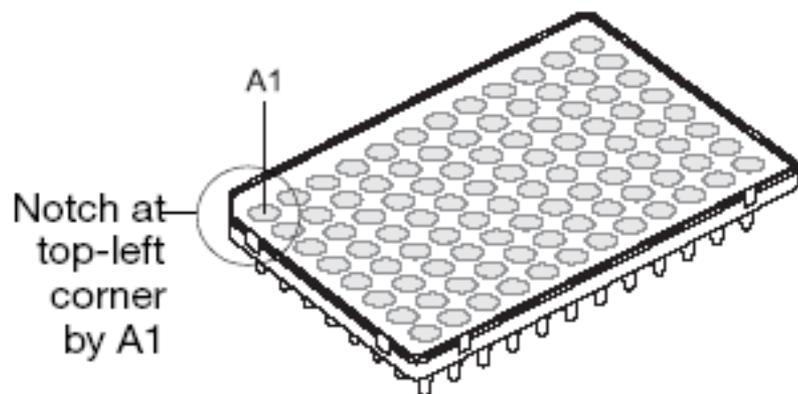
Zone 1	Zone 2	Zone 3	Zone 4
55.0 <input type="button" value="▲"/>	57.0 <input type="button" value="▲"/>	59.0 <input type="button" value="▲"/>	61.0 <input type="button" value="▲"/>



96 well

Fast 0.1 mL plates

Standard 0.2 mL plates



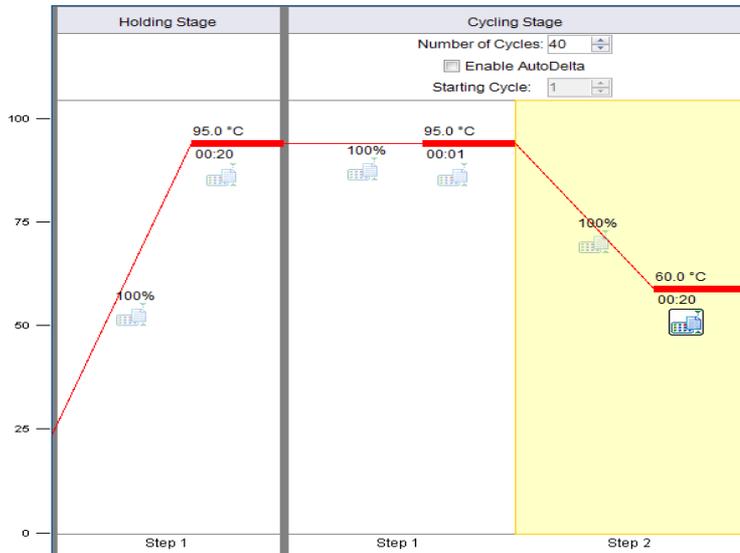
	<u>0.1mL</u>	<u>both</u>	<u>0.2mL</u>
96 well plates	4346906		4306737
96 well plate holder		4312063	
adhesive covers		4360954	
tray	4379983		4381850
8strip tubes	4358293		4316567
8strip tube caps		4323032	
Tube Capping tool		4330015	
single tubes	4358297		N8010540
Validated RXN volume	10-30uL		10-100uL

Instrument Default Cycling Options

Fast: 40 cycles in ~40min.

Standard: 40 cycles in ~1hr 45min

- faster ramping & shorter hold times



TaqMan Fast Advanced Master Mix (TaqPath ProAmp for SNP)

PowerUp SYBR Green Master Mix

There are many master mixes, read the protocol for the master mix!



123456789



Open/close

Set up run

Sean

QuantStudio® 5 - 96-Well 0.2-mL Block

Save the last 300 runs



My Account



Load Experiment



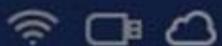
Open Template



Run Last



Settings



February 13, 2014 10:45am

Recommended Calibration and Verification

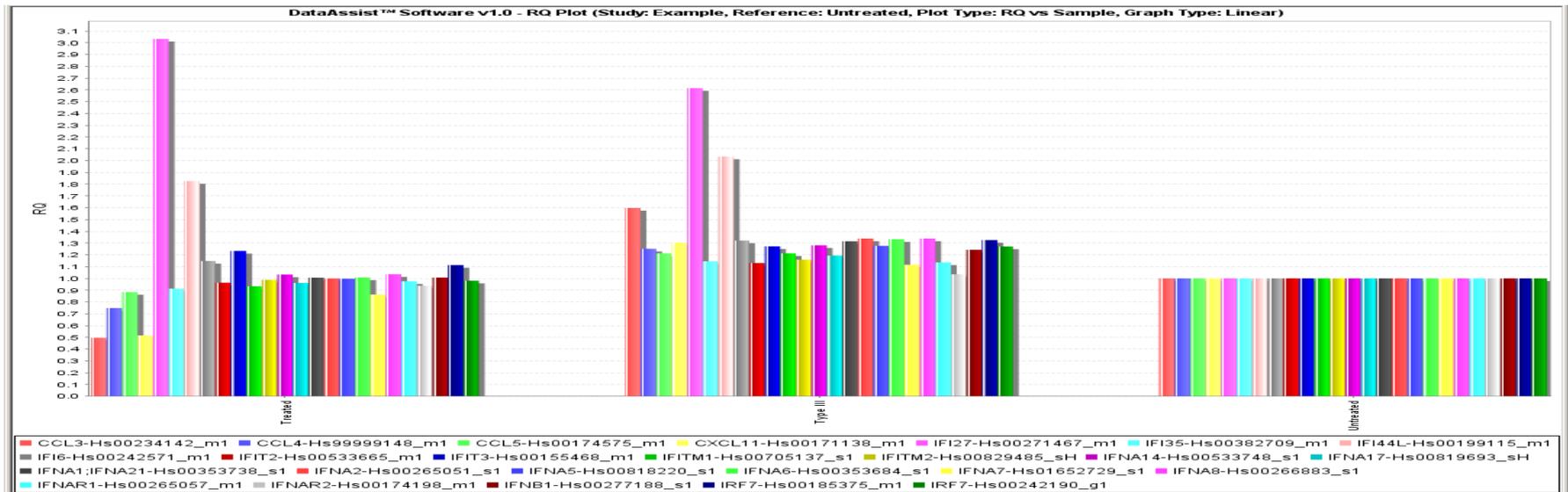
Calibration	Recommended frequency
ROI/uniformity	Every two years
	<ul style="list-style-type: none">Always perform new Background and Dye calibrations after an ROI/Uniformity calibration.
Dye	Every two years
	<ul style="list-style-type: none">During a Dye calibration, only the dyes on the given spectral calibration plate are calibrated.
Background	Every month
	<ul style="list-style-type: none">As needed: To check for contamination
	<ul style="list-style-type: none">As needed: To obtain the most accurate data for the removal of background fluorescence
RNase P verification	After installing or moving the instrument.
	After performing instrument or block calibrations.
	As needed to confirm instrument performance.

Thermo Fisher Connect

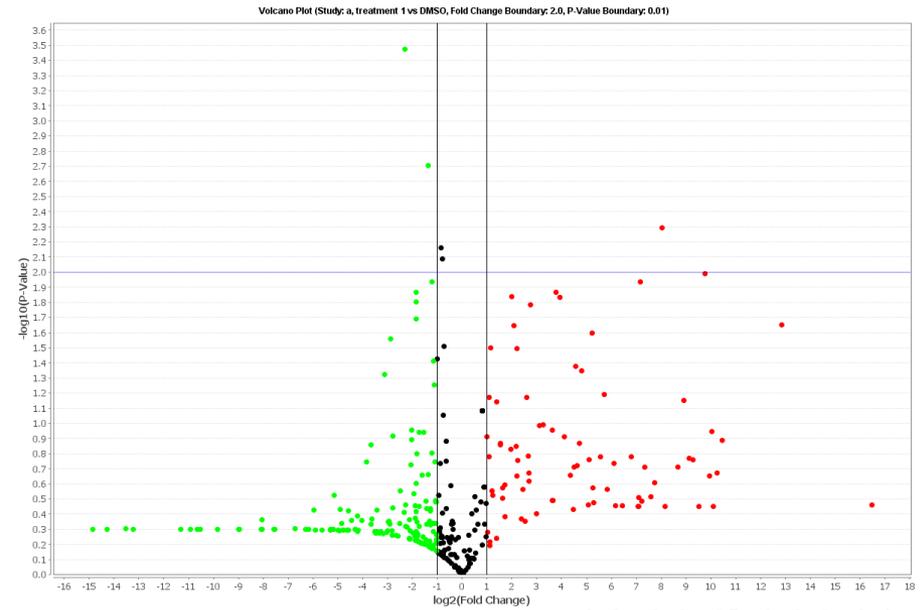
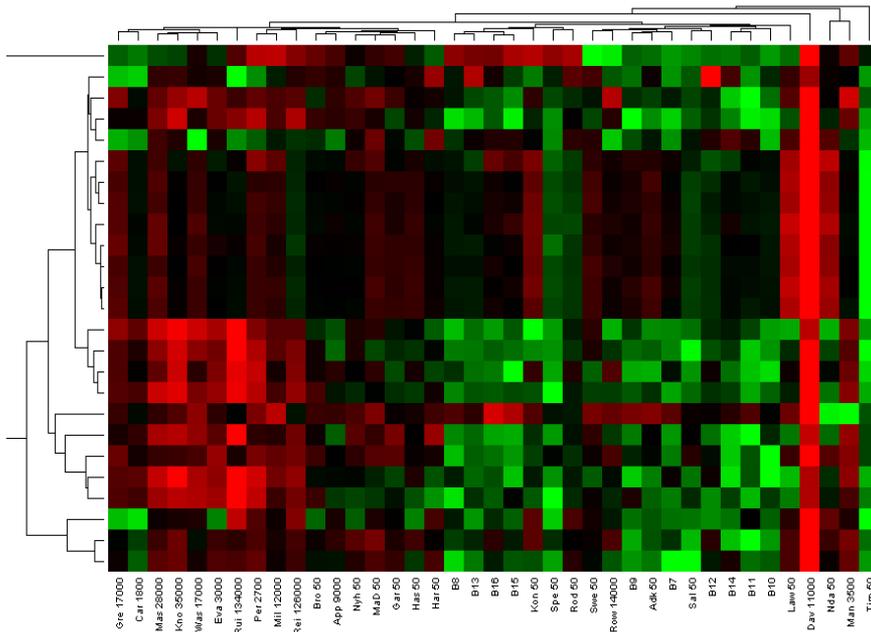
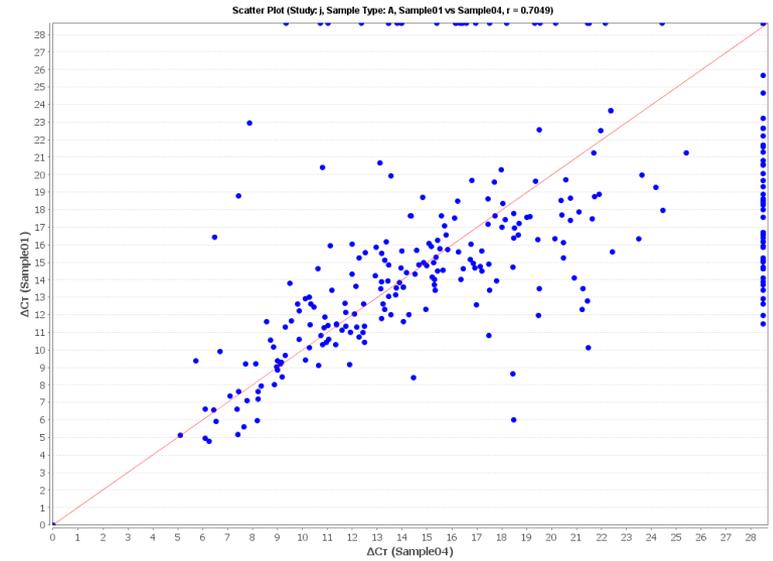
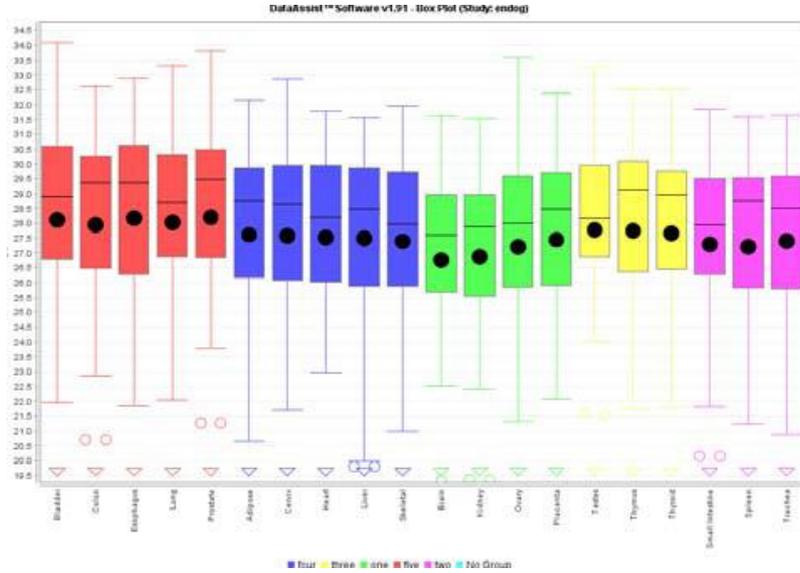
FREE web-based Gene Expression Analysis Software

How it works

- Import .eds data files from system software
- Analyze **>100 plates** in **one study**
- Graphical representation of the RQ data
- Export Analyzed Results, Data & Plots



Thermo Fisher Connect



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Rediscover your
scientific aspirations

aspire
member program



Join the free member program that supports your lifelong love of science.

Enroll today and receive a free, full-size product, 500 points towards rewards and discounts, and access to science career benefits and more...

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qPCR Applications & Information

Gene Expression

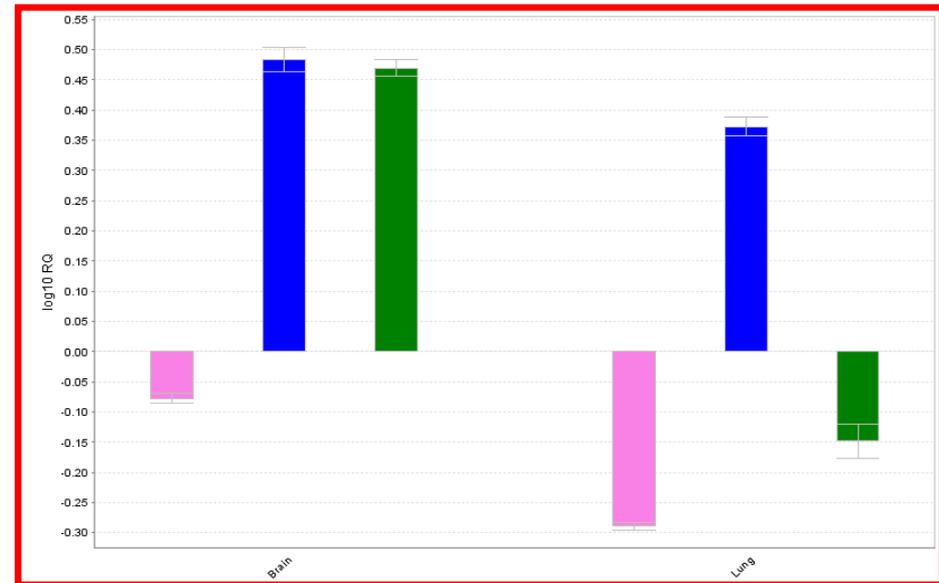
miRNA expression

SNP Genotyping

Copy Number Variation

CAST PCR for Mutation Analysis

Protein Thermal Shift



Digital PCR (QuantStudio 3D)
Service Plans on qPCR systems



Follow-up email with today's slides & survey

qPCR is the Technology to detect amplification of PCR products in real-time

Similar to traditional PCR...

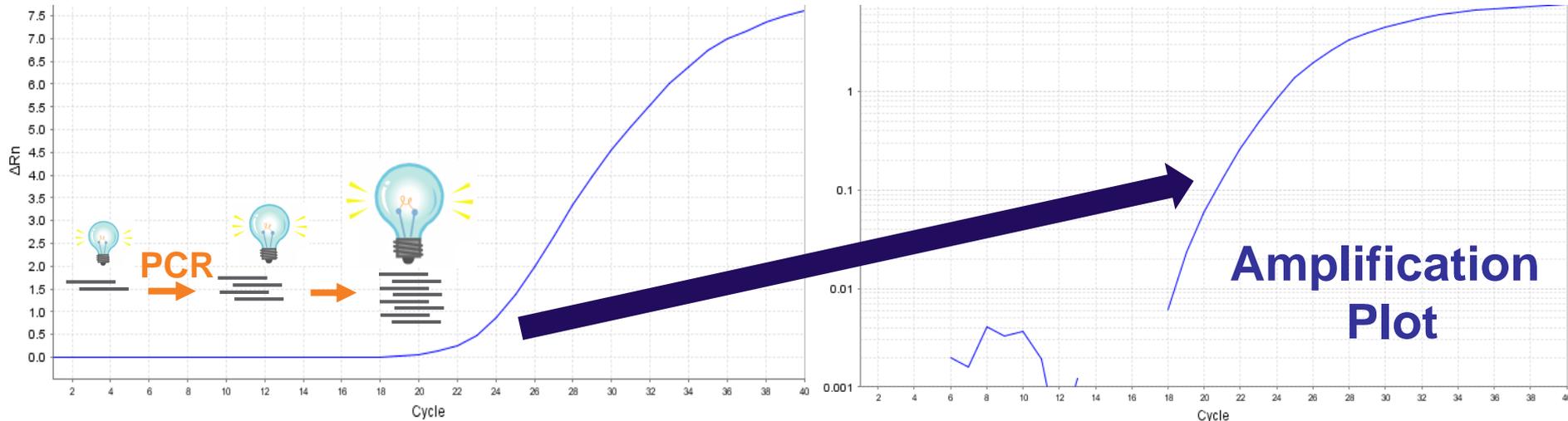


Denature template
Anneal primers
Extension of primers → amplicons

40 cycles

A **fluorescent** dye is in the reaction

As Amplification proceeds, fluorescence increases with PCR product formation



How to set up Real-Time PCR

Combine:

- 1) TaqMan® Master Mix or SYBR® Green Master Mix
- 2) TaqMan® Gene Expression Assay or SYBR® primers
- 3) water (#AM9935)
- 4) Sample (template)

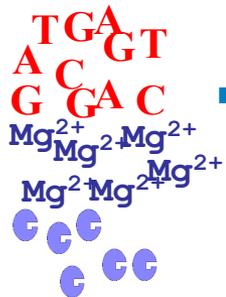
20uL reaction

(2X) 10uL

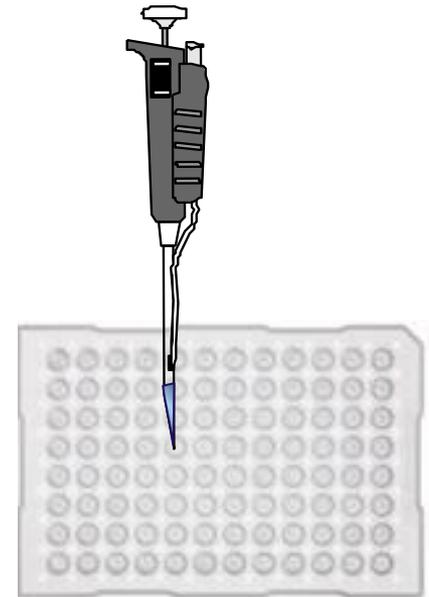
(20X) 1uL

7uL

(10 ng) 2uL



TGCAACAGATTAGACATAG
ATAGAGAATCTACAAACCGT



Nucleic Acid Isolation Kits

DNA & RNA Selection Guide
Displaying 7 results.

DNA RNA

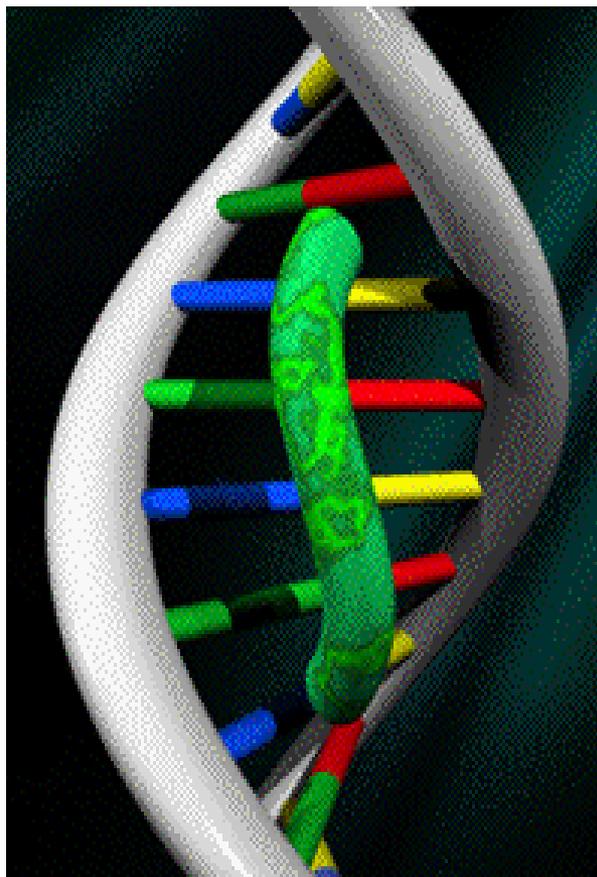
Sample Type	Starting Amount	Throughput	Application	Format
Cells	> less than 1000	> 1-50	> Real-Time	> Silica matrix column
Tissue	> 10^3 - 10^4	> 50-100	> RT-PCR	> Purification-free
Blood	> 10^4 - 10^5	> greater than 100	> microRNA	> Organic Solution
Virus	> greater than 10^5	>	> Cloning	> Magnetic beads
Bacteria/Yeast	>	>	> Microarrays	>
Plant	>	>	> Northern Blot	>
FFPE	>	>	> Sequencing	>
LCM	>	>	> messenger RNA	>

KingFisher Flex
Up to 96 samples in a run

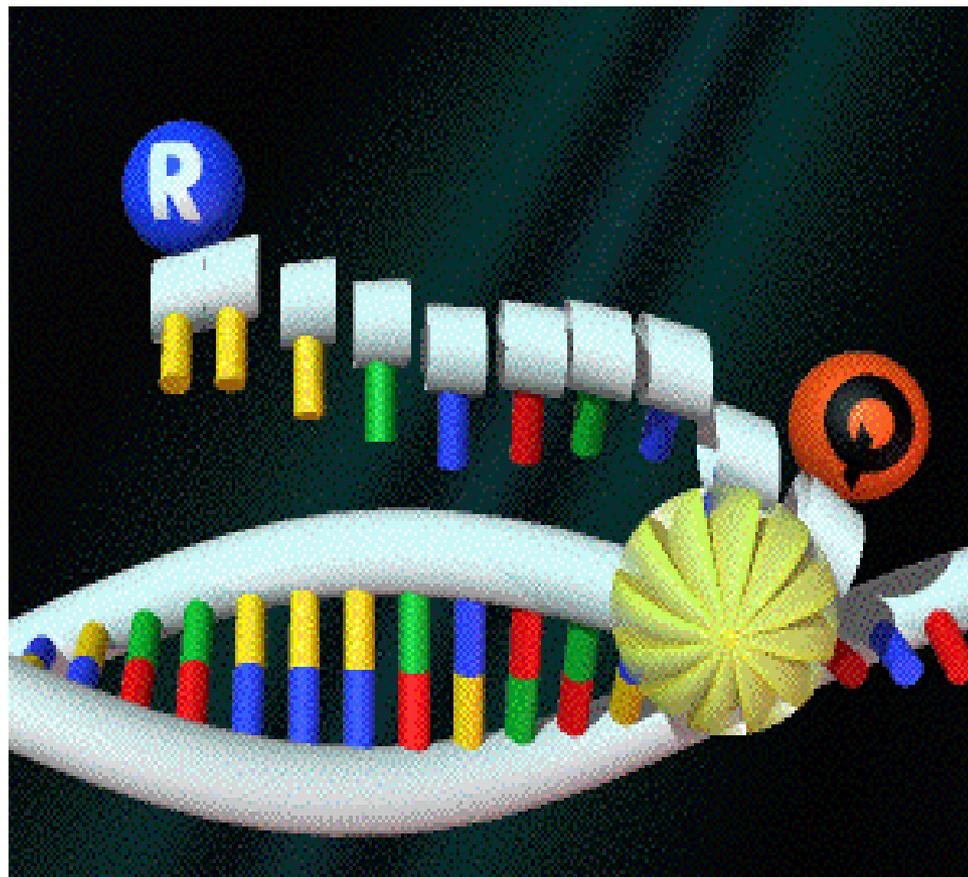


Supported Fluorescent Chemistries

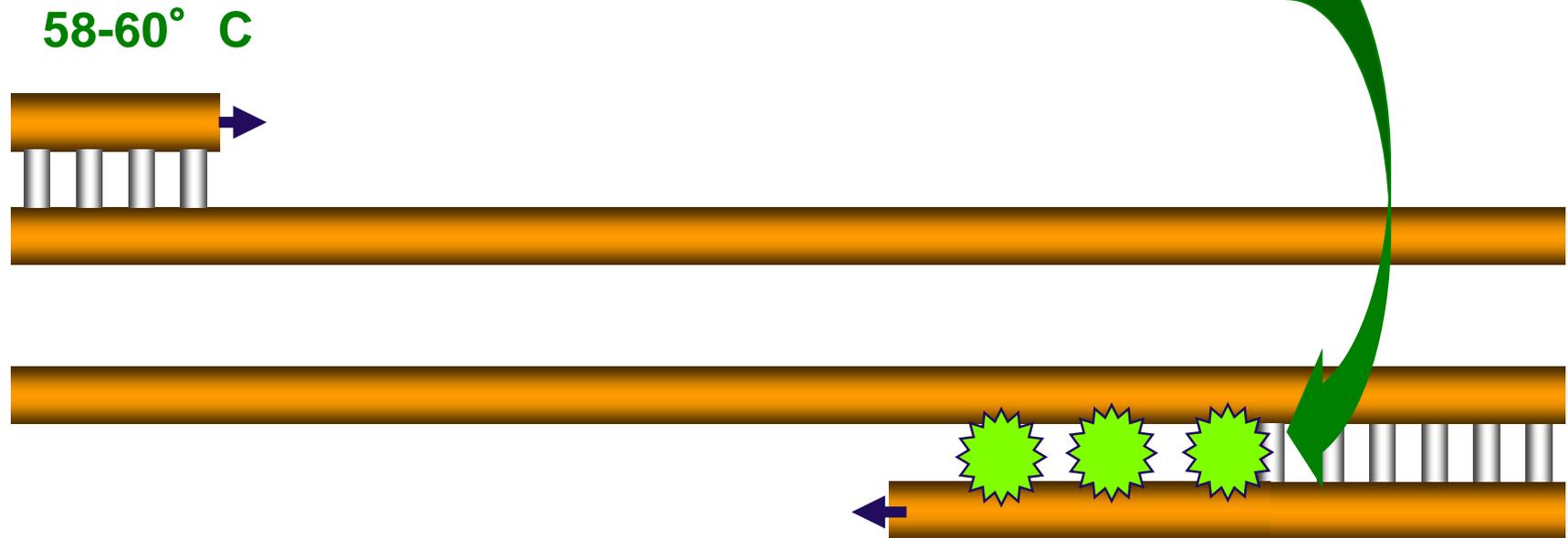
SYBR[®] Green



TaqMan[®]

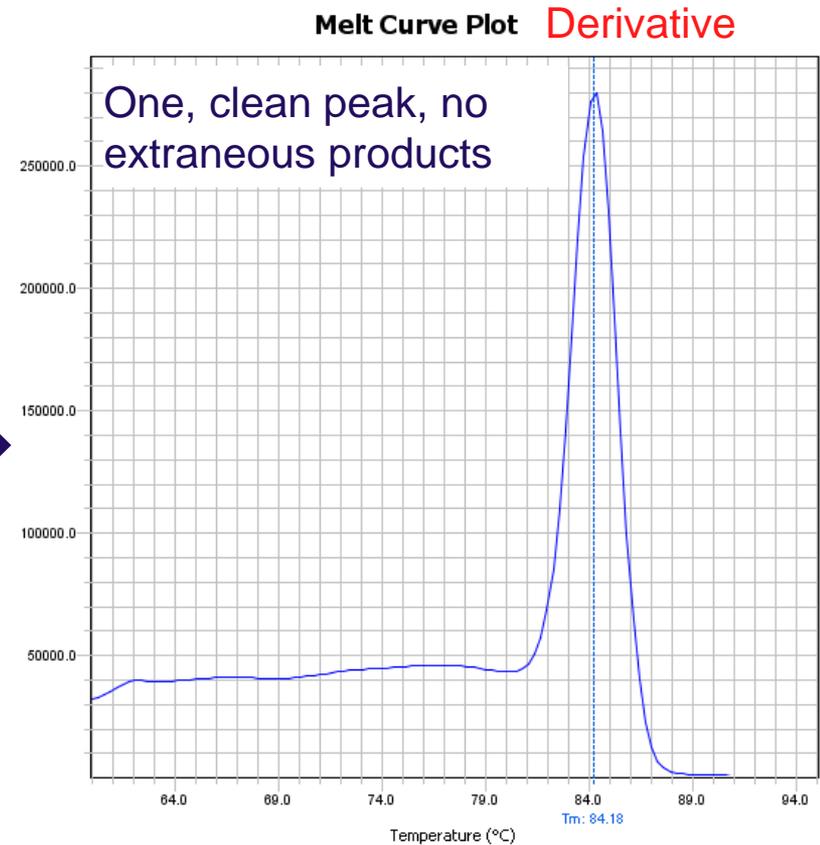
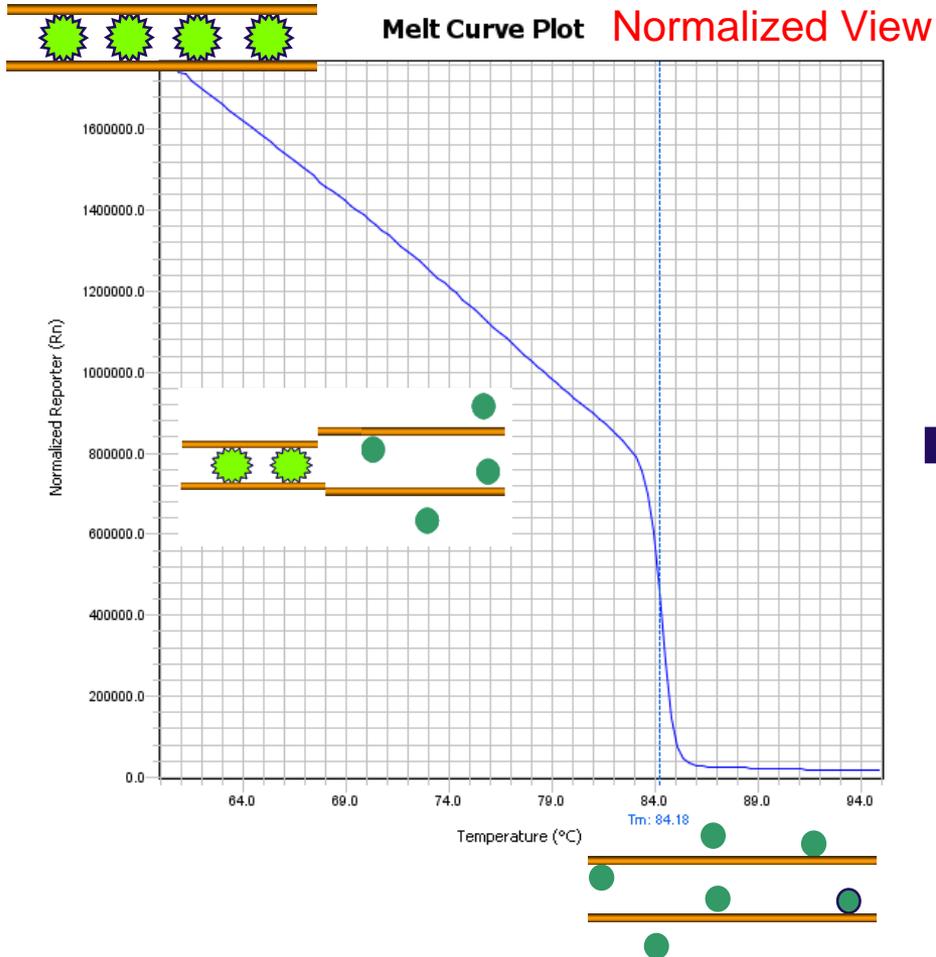


SYBR[®] Green Reaction

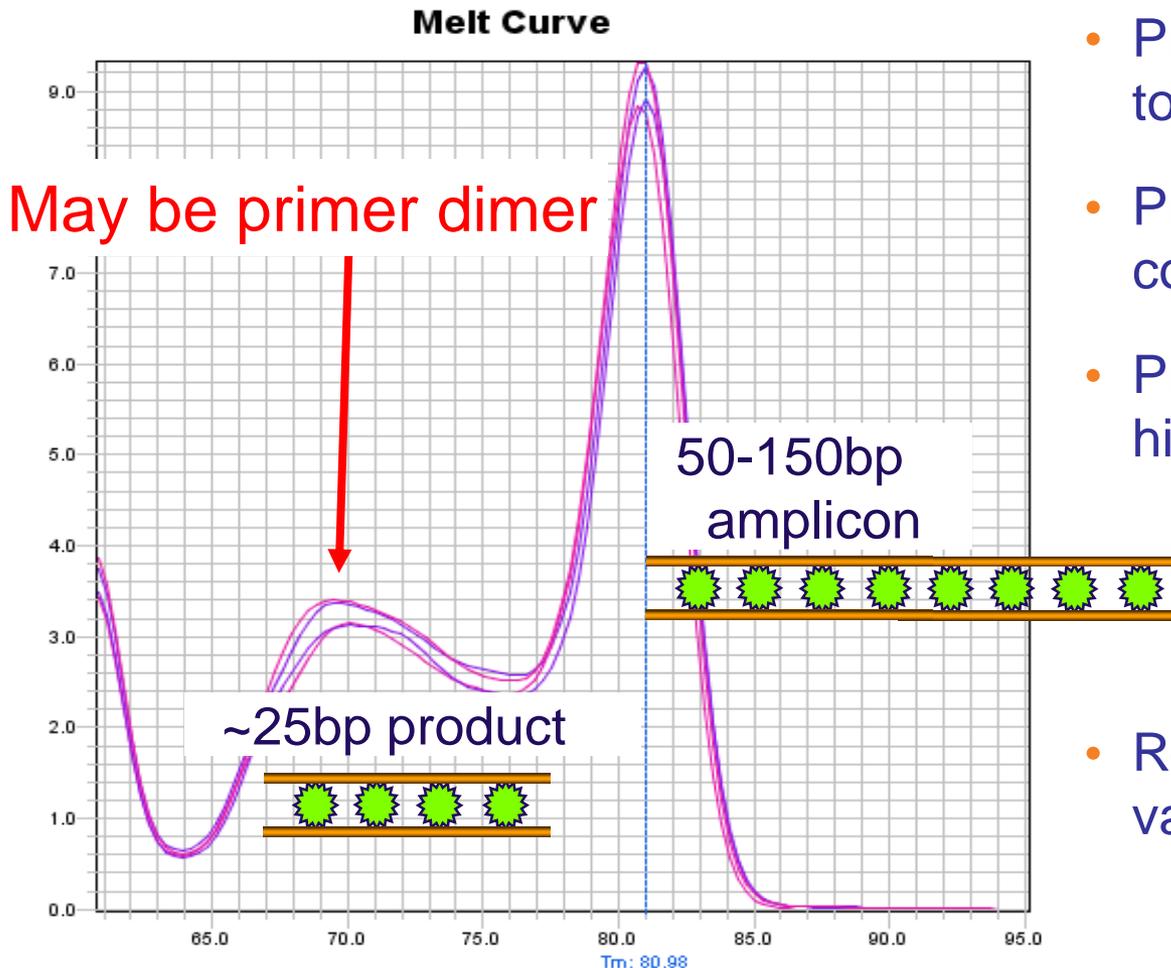


Binds to *any* double-stranded product in PCR reaction

Check specificity of reactions by Melt Curve protocol



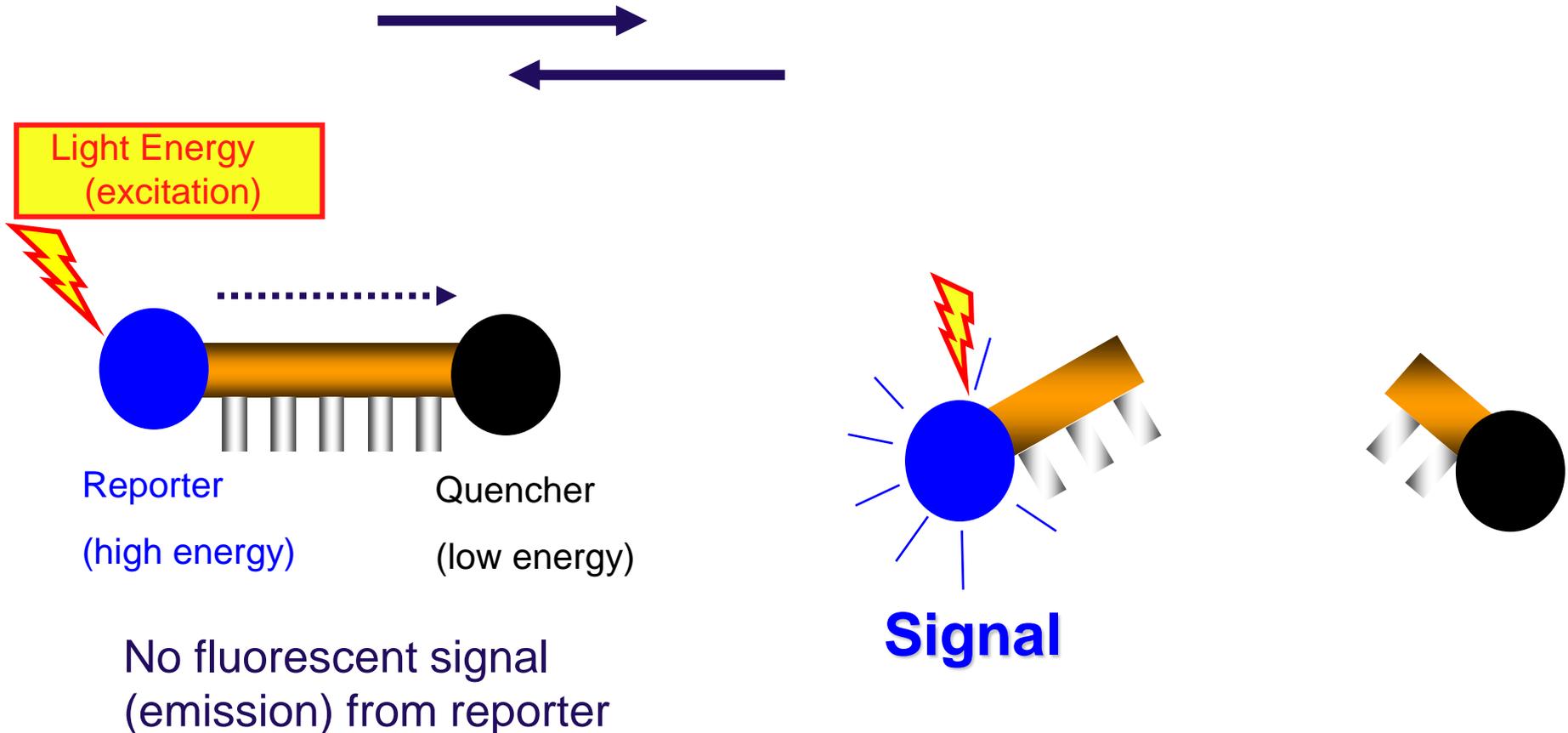
Extra peaks in melt curve



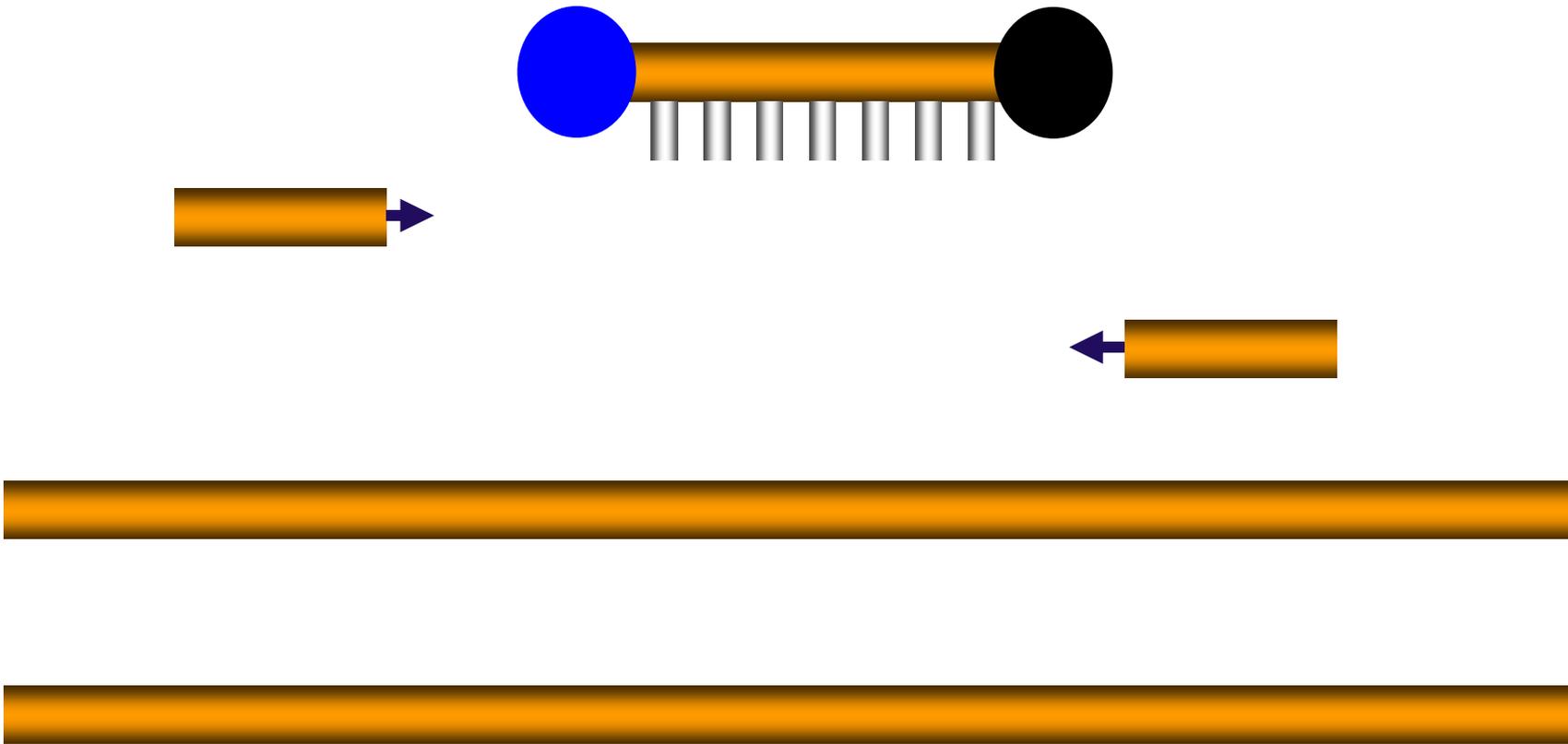
- Primer dimers can contribute towards fluorescent signal
- Primers are too complementary for each other
- Primer : template ratio is too high
- Resolved by testing out various primer concentrations

TaqMan Probe

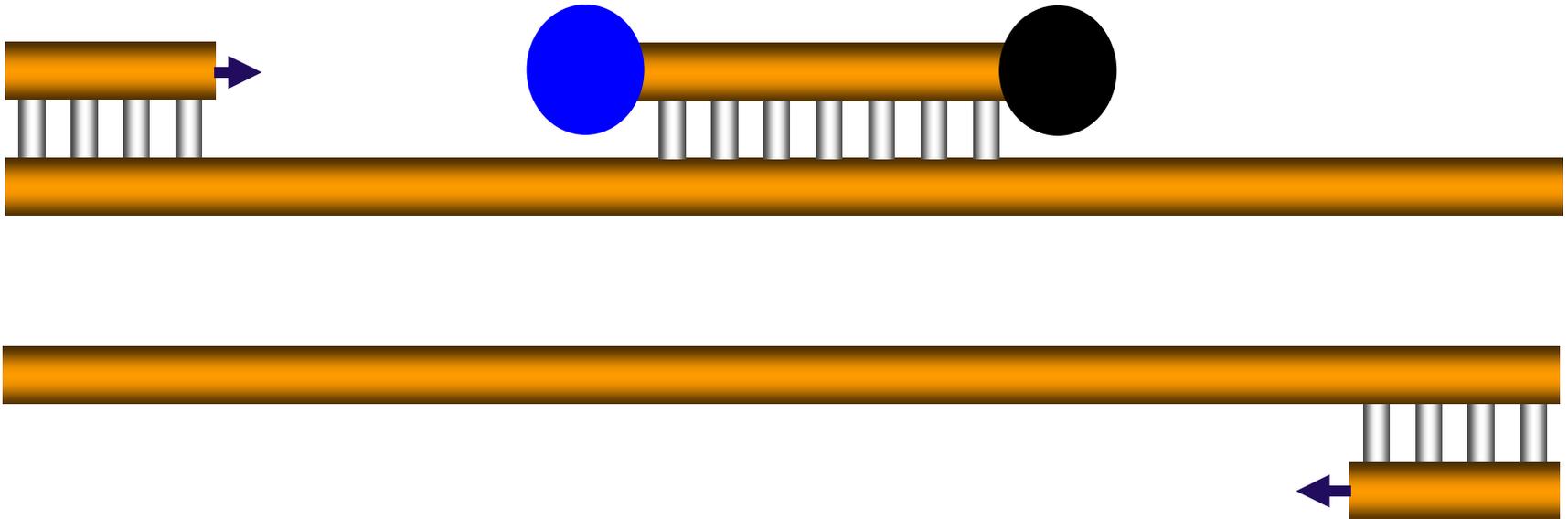
- **FRET**; phenomenon that describes an energy transfer mechanism between 2 fluorescent molecules when in close proximity.



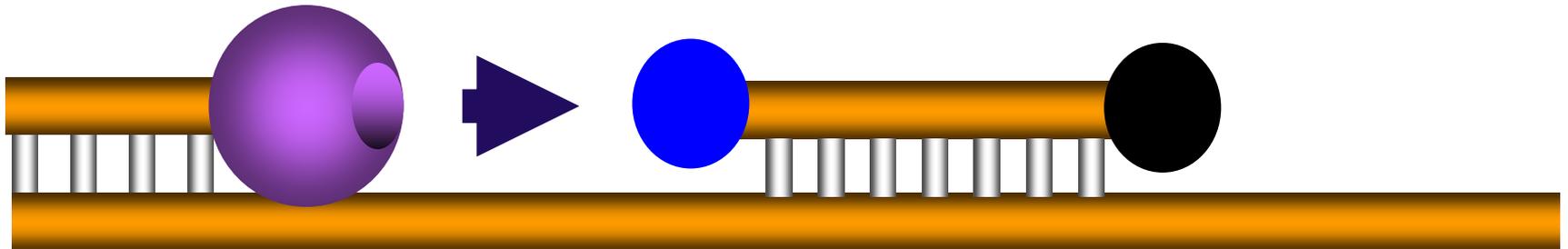
Denaturation (95°)



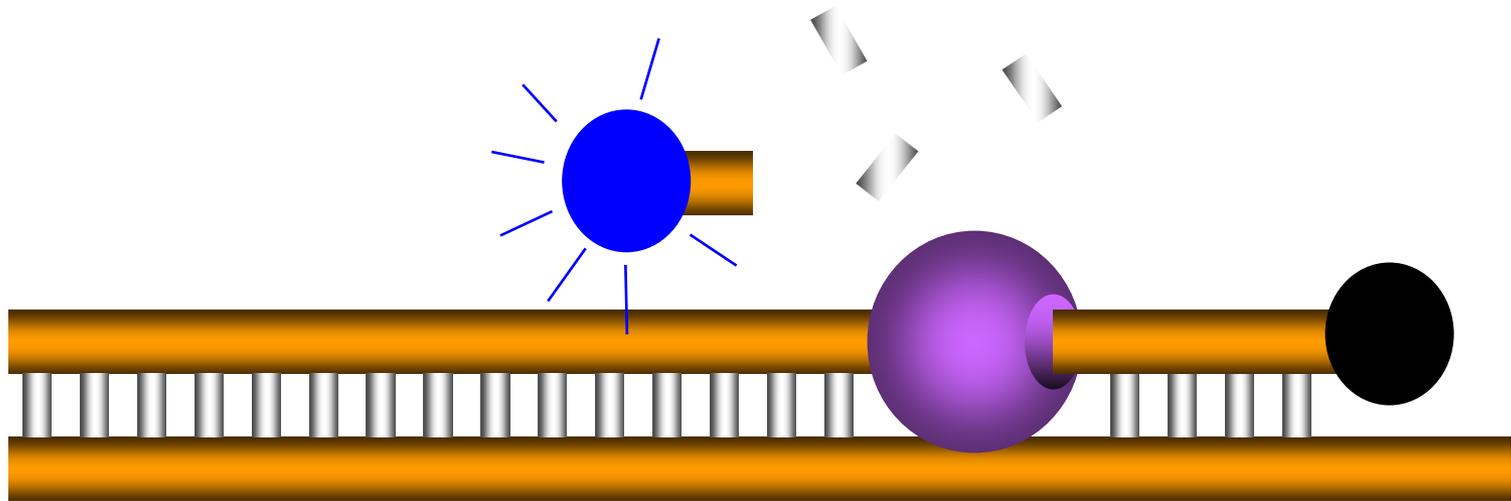
Annealing (60°)



Taq polymerase binds, then extends from upstream primer

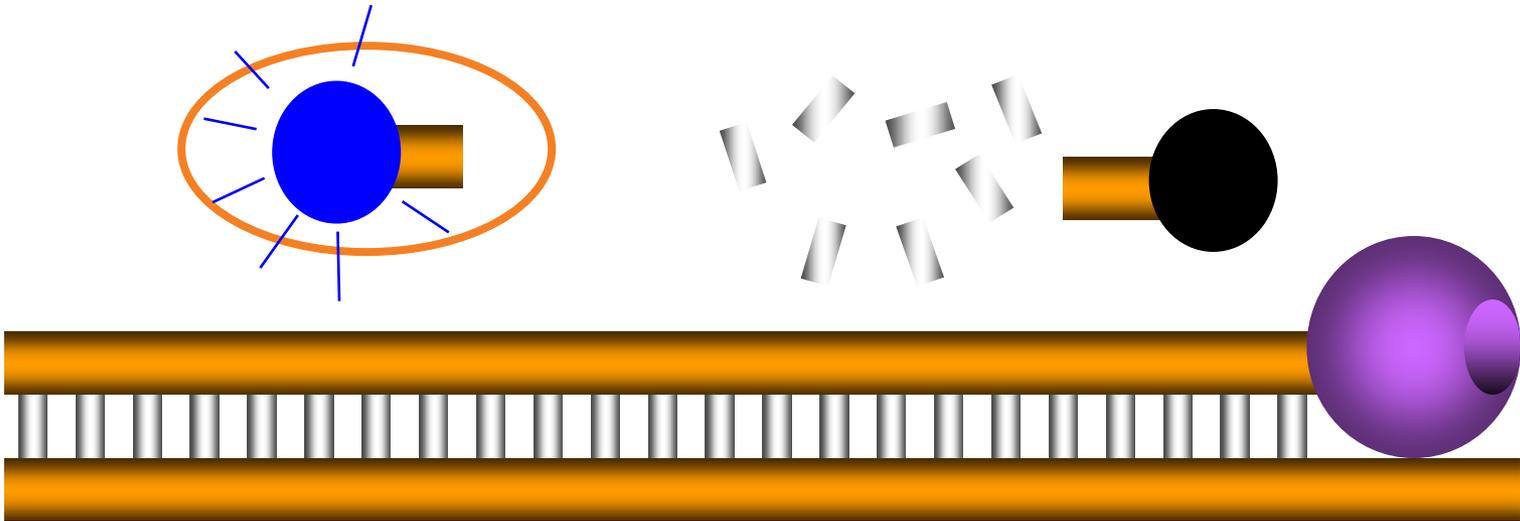


**Taq cleaves probe →
releases reporter dye**



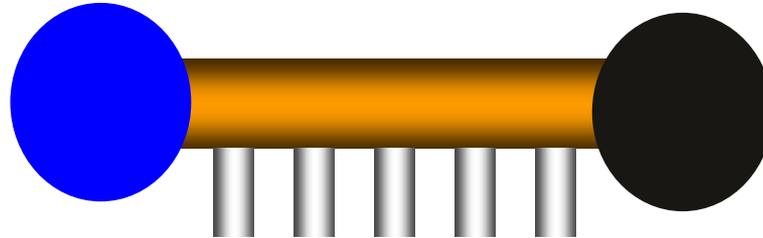
5'-Nuclease activity digests probe

Probe digested; *Taq* completes product



Reporter signal generated in tube/well

TaqMan[®] probe dye Choices



Reporter
(high energy)

Quencher
(low energy)

FAM[™]

VIC[™]

ABY[™]

JUN[™]

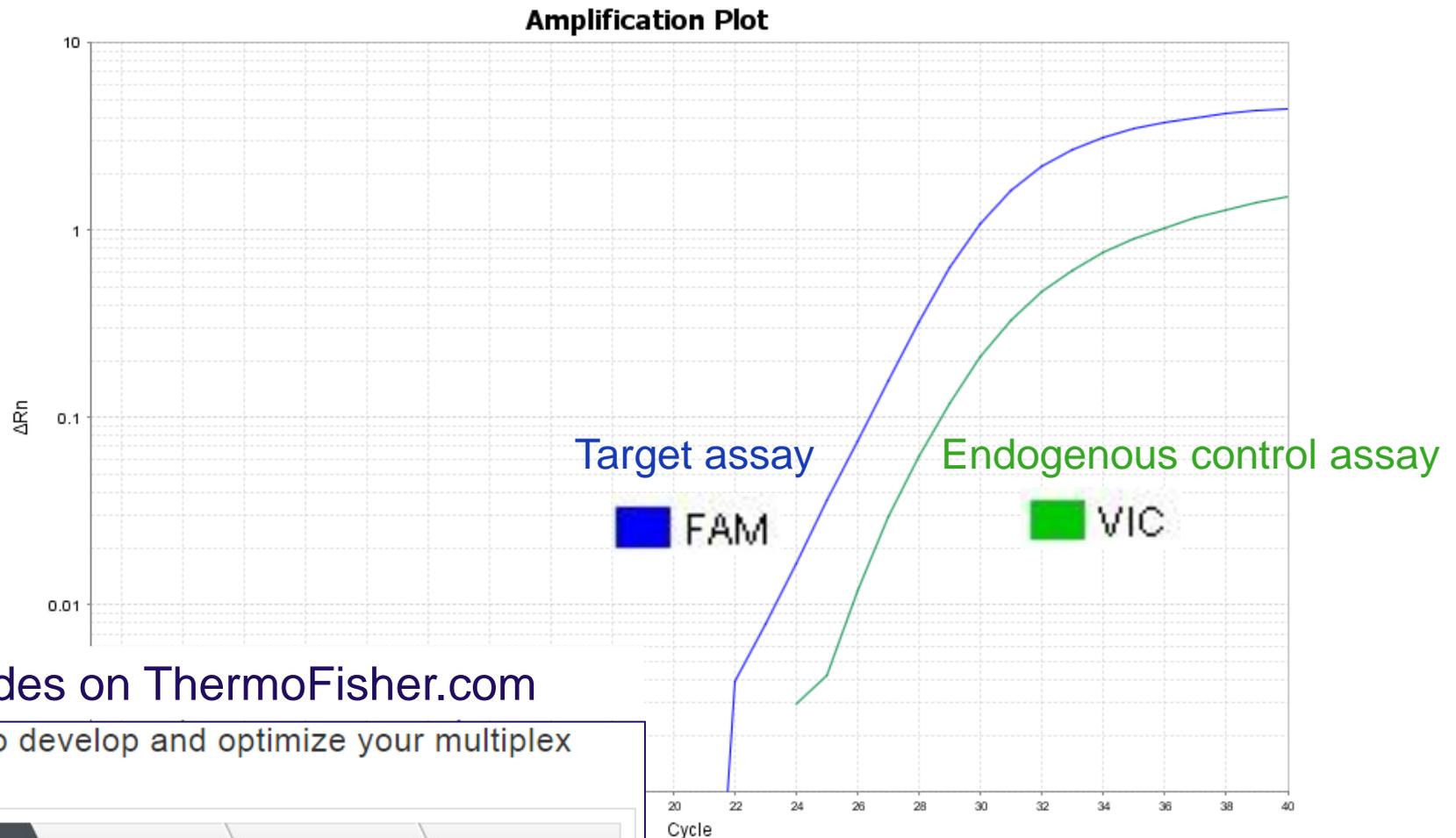
MGB (non-fluorescent)

TAMRA[™]

QSY - for multiplexing >2 targets

- easily convert from BHQ

Pure Dye Spectra of System Enables Multiplexing (TaqMan chemistry)



guides on [ThermoFisher.com](https://www.thermofisher.com)

Workflow to develop and optimize your multiplex reaction

Step 1

Determine your probe options

Step 2

Redesign and order assays, primers, and probes

Step 3

Choose and order the appropriate master mix

Step 4

Optimization

Assay Set-up

SYBR

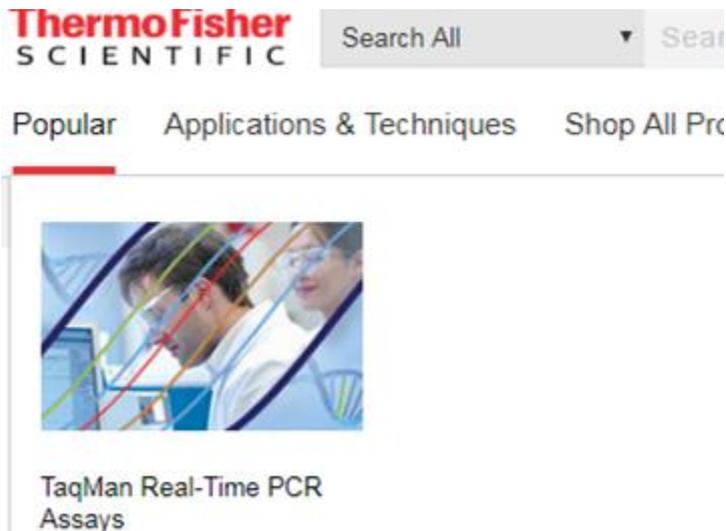
- primer optimization experiment... 300nM, 200nM, 100nM
- Use the concentration that provides the strongest amplification fluorescent signal and a single product in the melt curve

TaqMan

- Primers @ **900nM**, Probe @ **250nM** (final concentration in reaction).
- Probe T_m (**68-70c**) is ~10c higher than primers (**58-60c**)
- **Pre-designed assays (20X no need to worry about this!)**



TaqMan[®] Gene Expression Assays



The screenshot shows the ThermoFisher Scientific website interface. At the top left is the logo. To its right is a search bar with the text 'Search All' and a dropdown arrow. Below the search bar are navigation links: 'Popular', 'Applications & Techniques', and 'Shop All Products'. A large image shows two scientists in a lab setting with colorful lines overlaid. Below the image is the text 'TaqMan Real-Time PCR Assays'.

Human	Soybean	Wine grape
Mouse	Rhesus Monkey	Clawed frog
Rat	Rabbit	Chicken
Arabidopsis	Rice	Wheat
Cattle	Pathogen	Frog
Nematode	Baker's yeast	Chinese hamster
Dog	Fission yeast	Maize
Guinea Pig	Pig	Fruit fly
Zebrafish	Bread wheat	Corn
Horse		

2,300,000
Pre-developed Assays

1 tube contains; Primers & probe (Optimized at 20x concentration)

“Probe spans Exons”: no gDNA detected

Popular Applications



TaqMan Real-Time PCR Assays

Hs02786624_g1 GAPDH 4 RefSeq (NM) Both primers and probe 157 Dye: FAM-MGB

Catalog n
Target sp
Importan

View Details ▾ Re

99 / 100
Bioss Stars Ⓞ

View Details ▾ Re

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Bioss Stars Ⓞ

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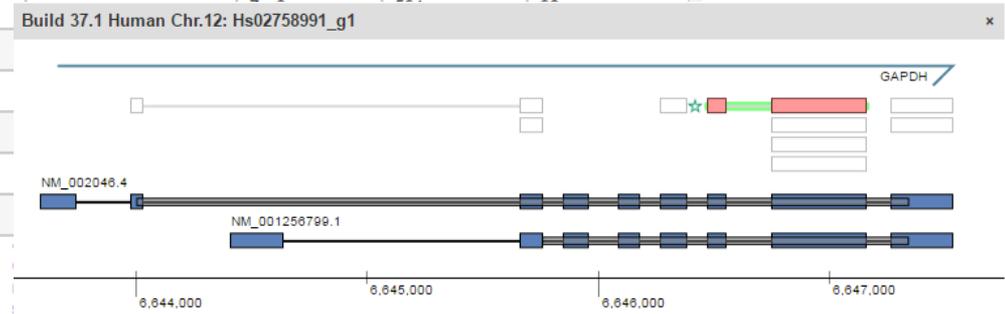
View Details ▾ Re

99 / 100
Bioss Stars Ⓞ

View Details ▾ Re

Gene Transcripts
Gene Symbol: GAPDH
Entrez Gene ID: [2597](#)
Gene Name: glyceraldehyde-3-phosphate dehydrogenase
Gene Aliases: CDABP0047, G3PD, GAPD
Location Chromosome: Chr.12: 6643657 - 6647536 on Build 37
UniGene: [Hs.544577](#)

Interrogated Sequence		Translated Protein	Exon Boundary	Assay Location	Amplicon Length
RefSeq	NM_001256799.1	NP_001243728.1	6 - 7	728	93
	NM_002046.4	NP_002037.2	7 - 8	704	93
GenBank mRNA	AB062273.1	-	7 - 8	582	93
	AF261085.1	-	7 - 8	633	93
	AK026525.1	-	7 - 8	655	93
	AK308198.1	-	7 - 8	606	93
	AY007133.1	-	7 - 8	582	93
	AY633612.1	-	6 - 7	554	93
	BC001601.1	-	7 - 8	584	93
	BC004109.2	-	7 - 8	582	93
	BC009081.1	-	7 - 8	582	93
	BC013310.2	-	7 - 8	582	93



Availability: **Inventoried**
Price (USD): **173.00**
[Check your price](#)

Add to cart

Connect Your Lab



Cell Culture Plas

Custom TaqMan[®] Assay Design Tool

Status	Assay Name	Primer Sequence	Probe Sequence
	<input type="text"/>	Forward Primer: <input type="text"/> Reverse Primer: <input type="text"/>	Probe 1 Dye: <input type="text" value="6-FAM"/> <input type="text"/> <input type="text"/> Do you wish to permanently hide your assay primer/probe sequences in all documents? More Info <input checked="" type="radio"/> No <input type="radio"/> Yes

Status	Name	Sequence	Target Position & Name
Valid	<input type="text" value="TEST"/>	GTCAGAGAGAAAGTAGGAGGCCATTGAAGTCGTCGGACTGAGGAACAA AATAACACAAGTGTCTAGAGAGAAAGTAGGAGGCCATTGAAGTCGTCGG ACTGAGGAACAAAATAACACAAGTGTCTAGAGAGAAAGTAGGAGGCCATT GACTCTCCGACATCAAGCAAGTAAGCAAGTGTCTAGAGAGAAAGTAGGAGGCCATT Do you wish to permanently hide your assay primer/probe sequences in all documents? More Info <input checked="" type="radio"/> No <input type="radio"/> Yes	<input type="radio"/> Manual <input checked="" type="radio"/> Automatic <input type="text" value="ANY"/> <input type="text" value="ANY"/>



Design Job Details

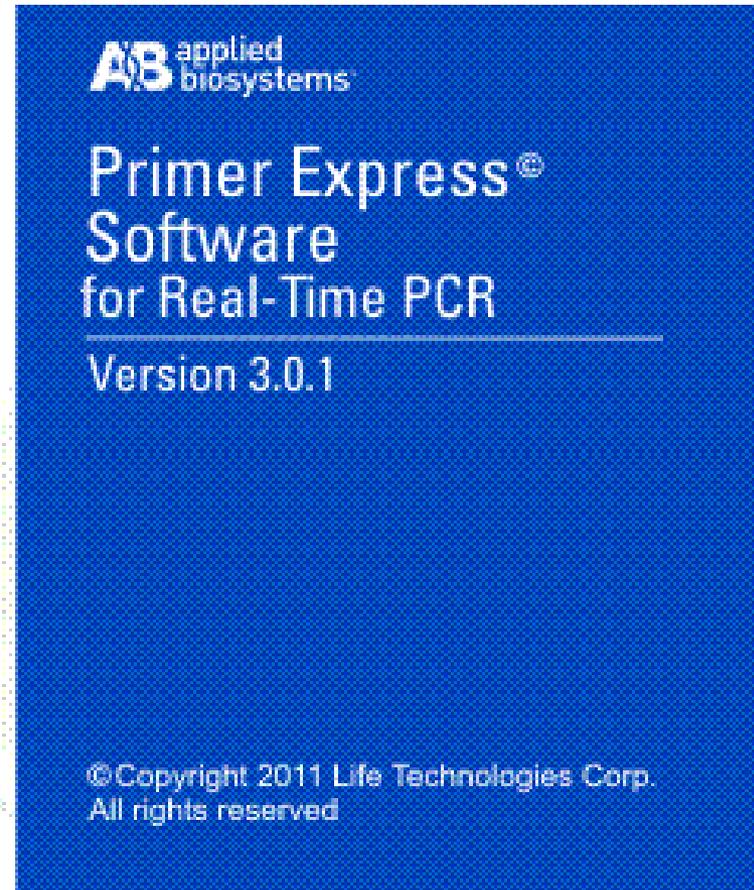
Refresh Batch List

Batch ID	Submitted	Status	Details
w0906118406000	2009-06-13 16:59:41	COMPLETED	1 Passed, 0 Failed, 0 No_Design

Design Results

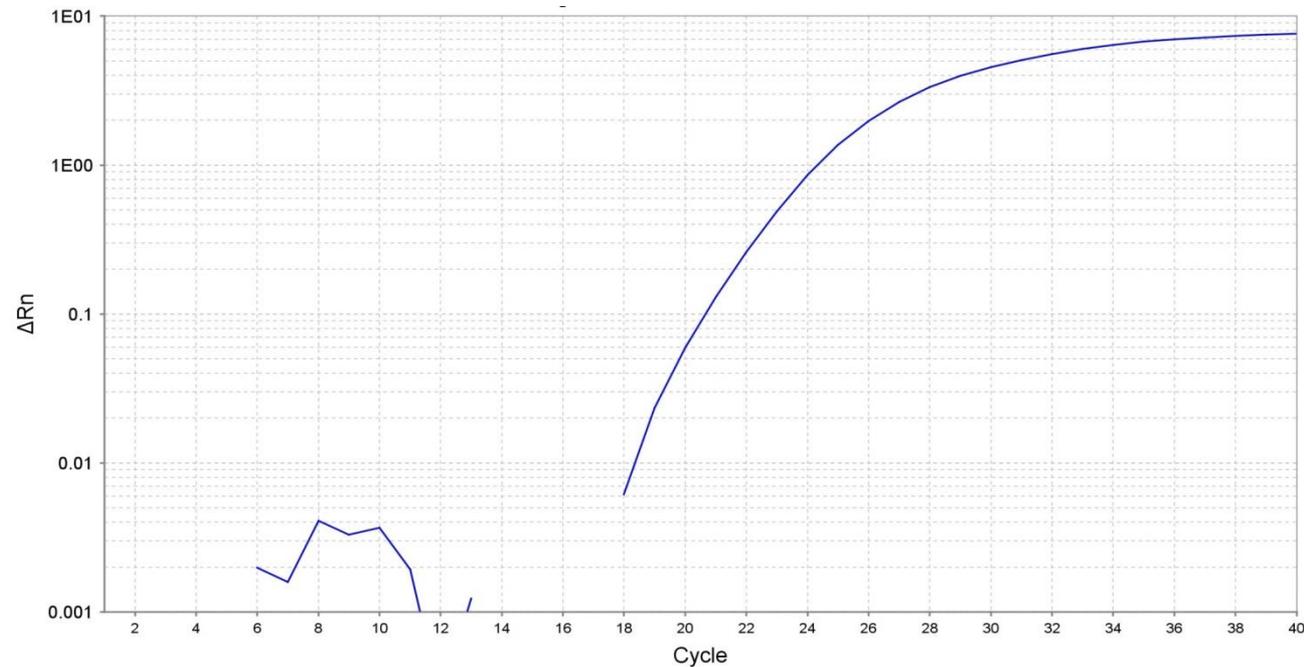
ID / Name	Type	Design Status	Size	Quantity	Add All
AILIWHU TEST	Custom	Passed	4331348 : small	<input type="text" value="1"/>	<input type="button" value="Add"/>

Primer Express® Software



<https://www.thermofisher.com/order/catalog/product/4363991>

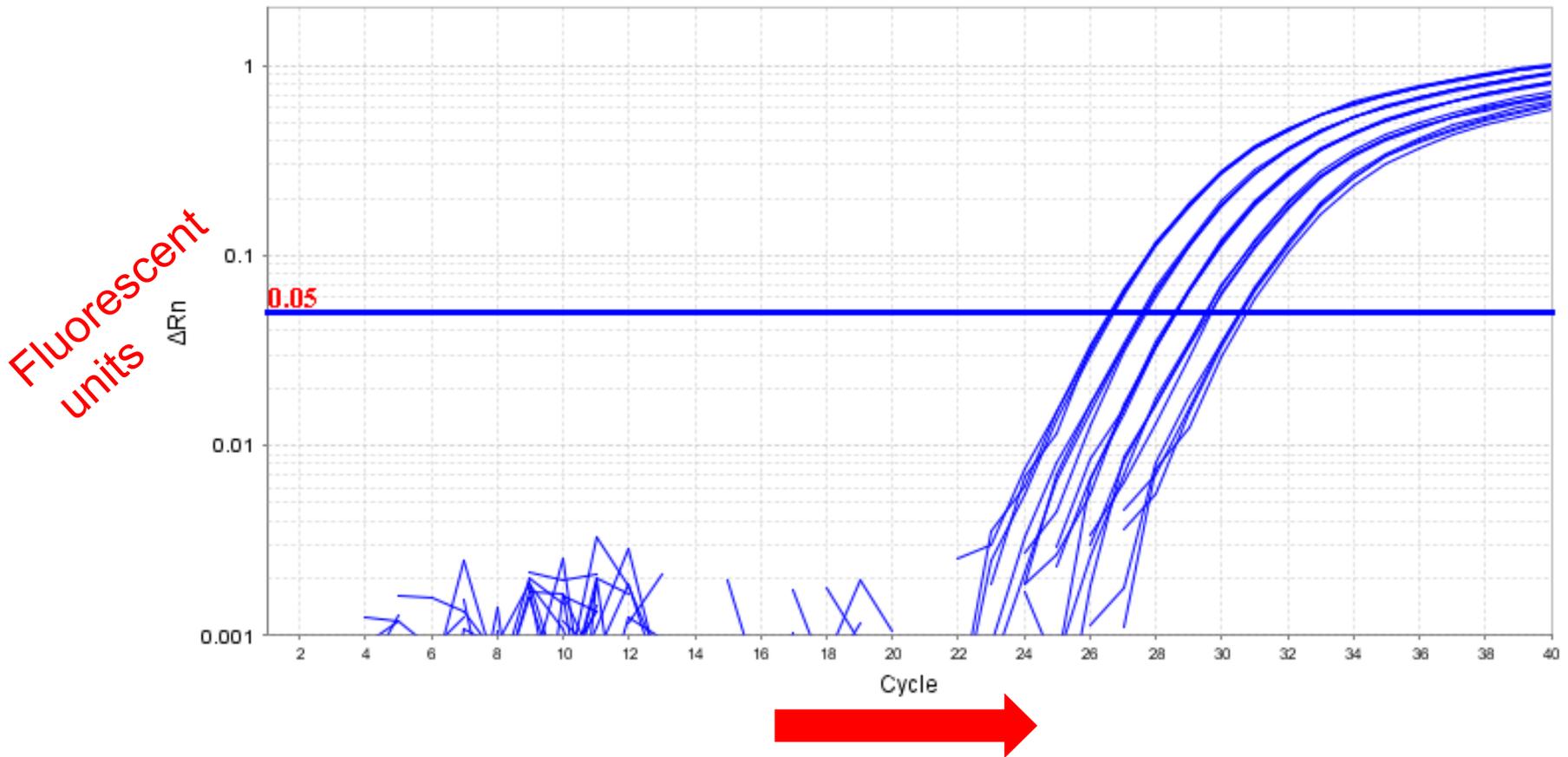
Examining qPCR amplification plots



Real-time PCR Terminology

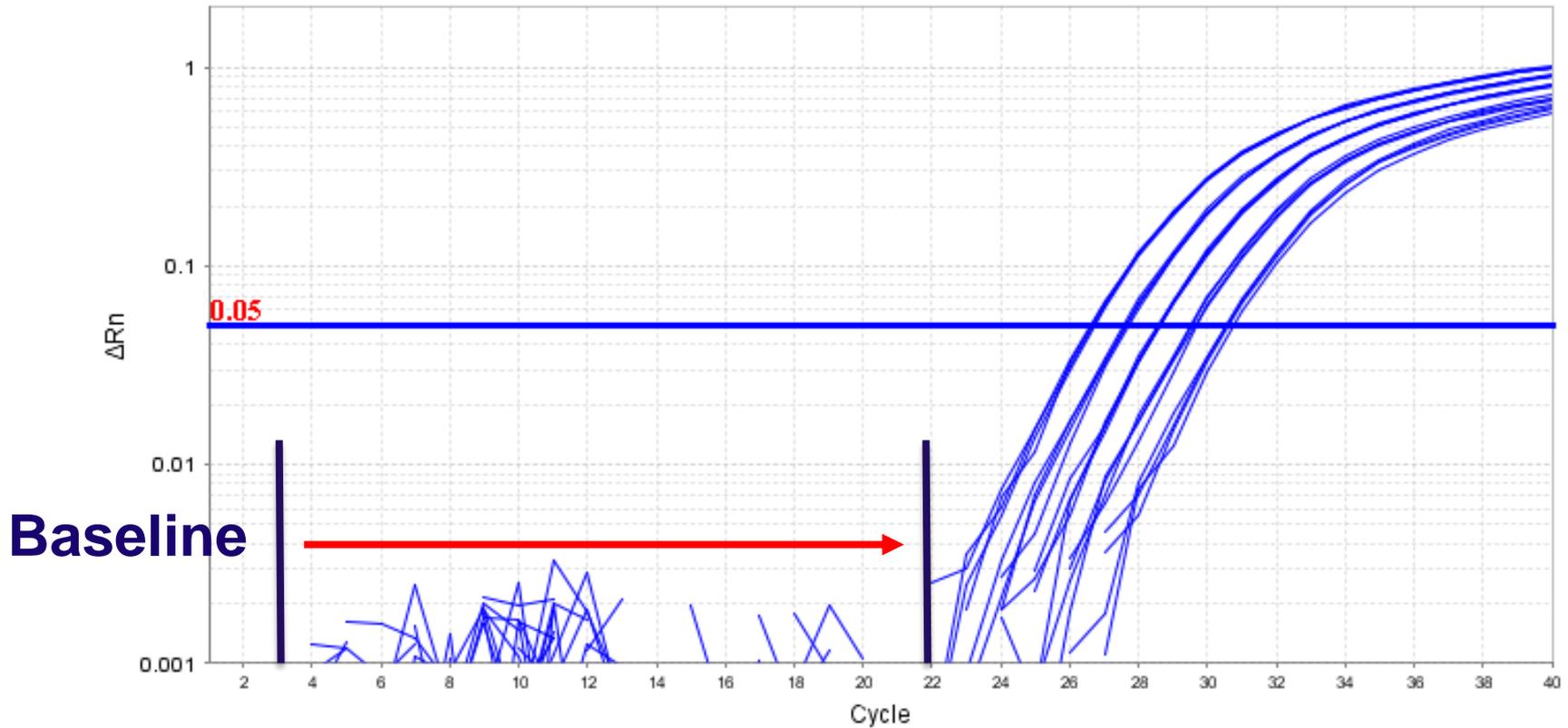
Amplification plots

graph cycle vs. Δ in fluorescence



Baseline

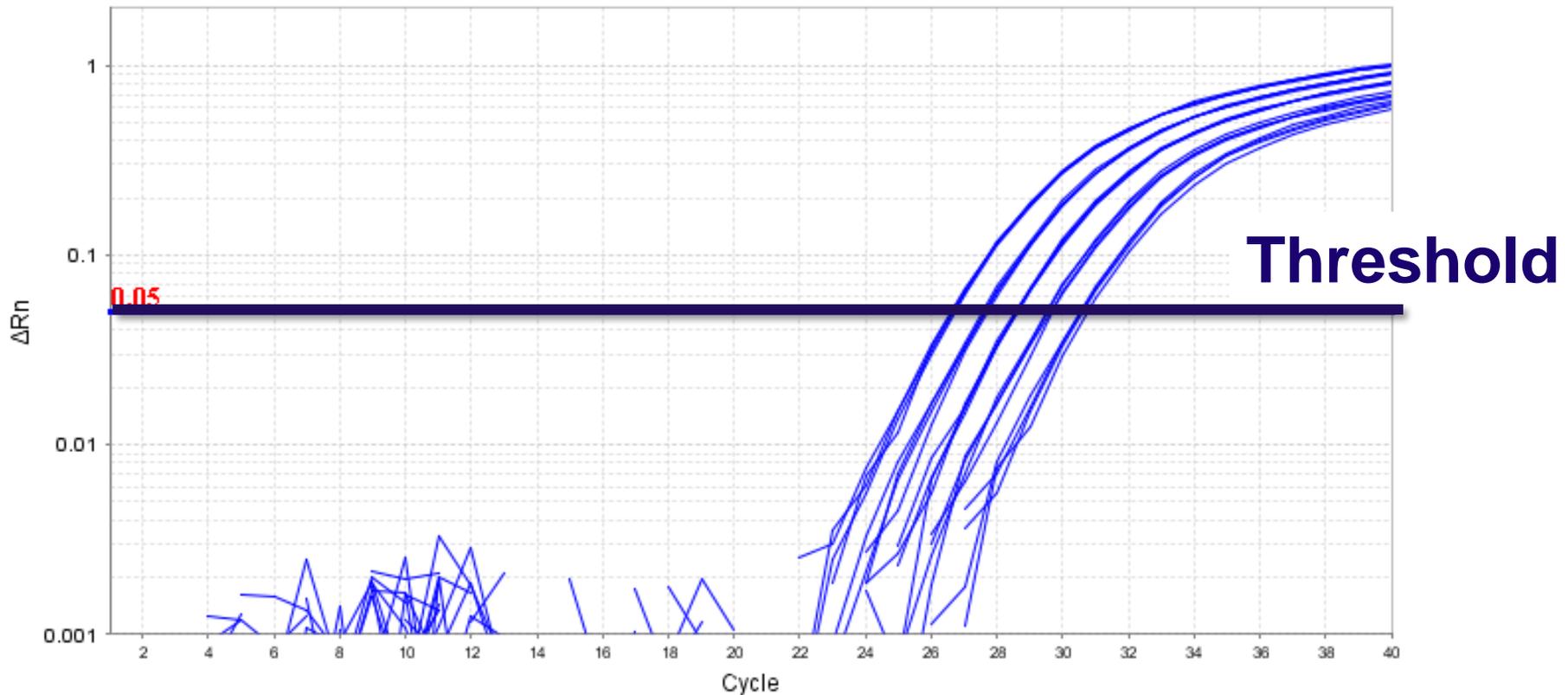
Defines region of background noise by a cycle-to-cycle range ex. cycles 3-22



Automatic/manual baseline

Threshold

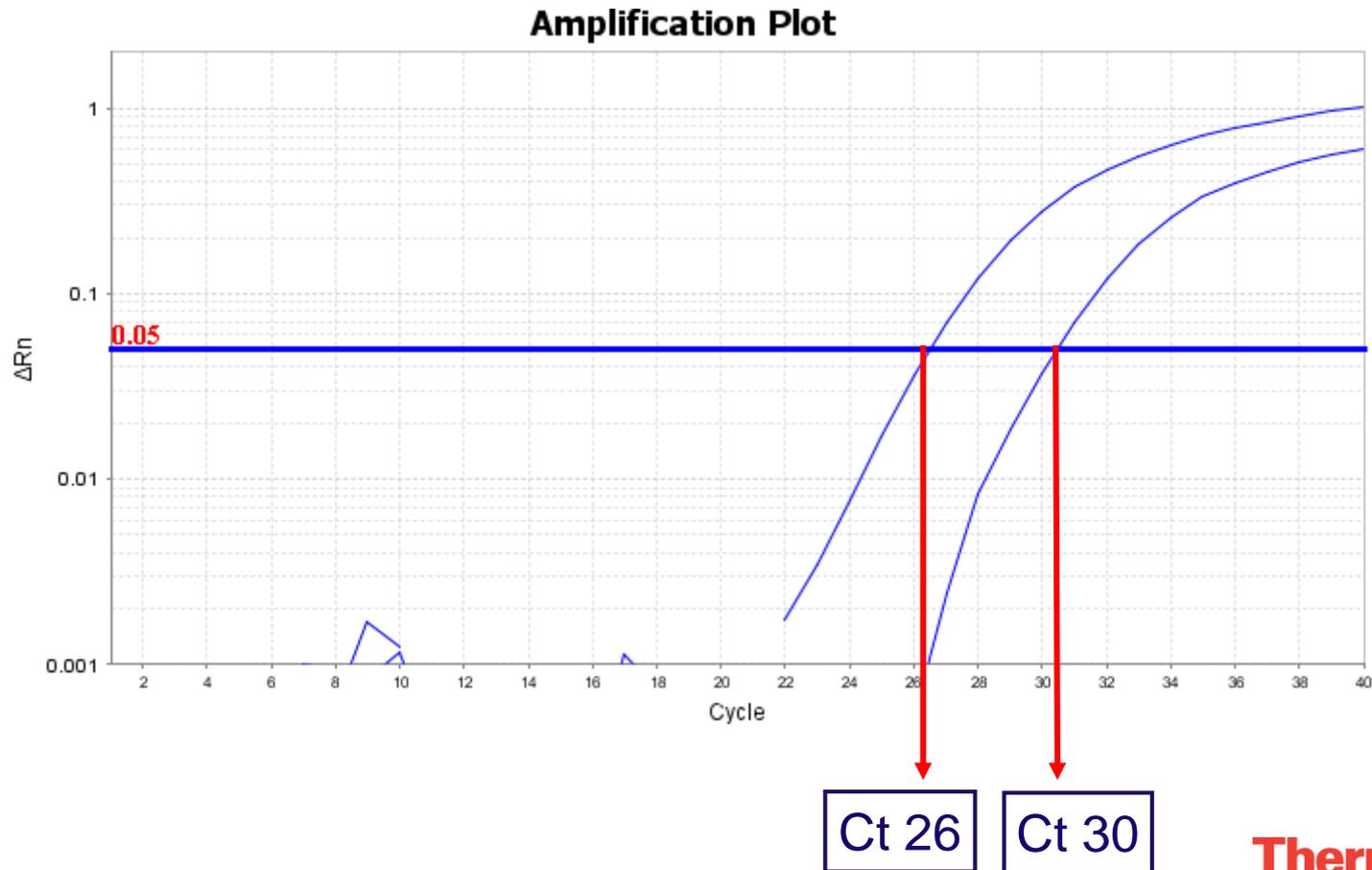
An adjustable horizontal line in the amplification plot.
Tells the software where to capture the data.



AutoThreshold (default); Software sets the threshold for each assay.
Different assays may have different exponential phases

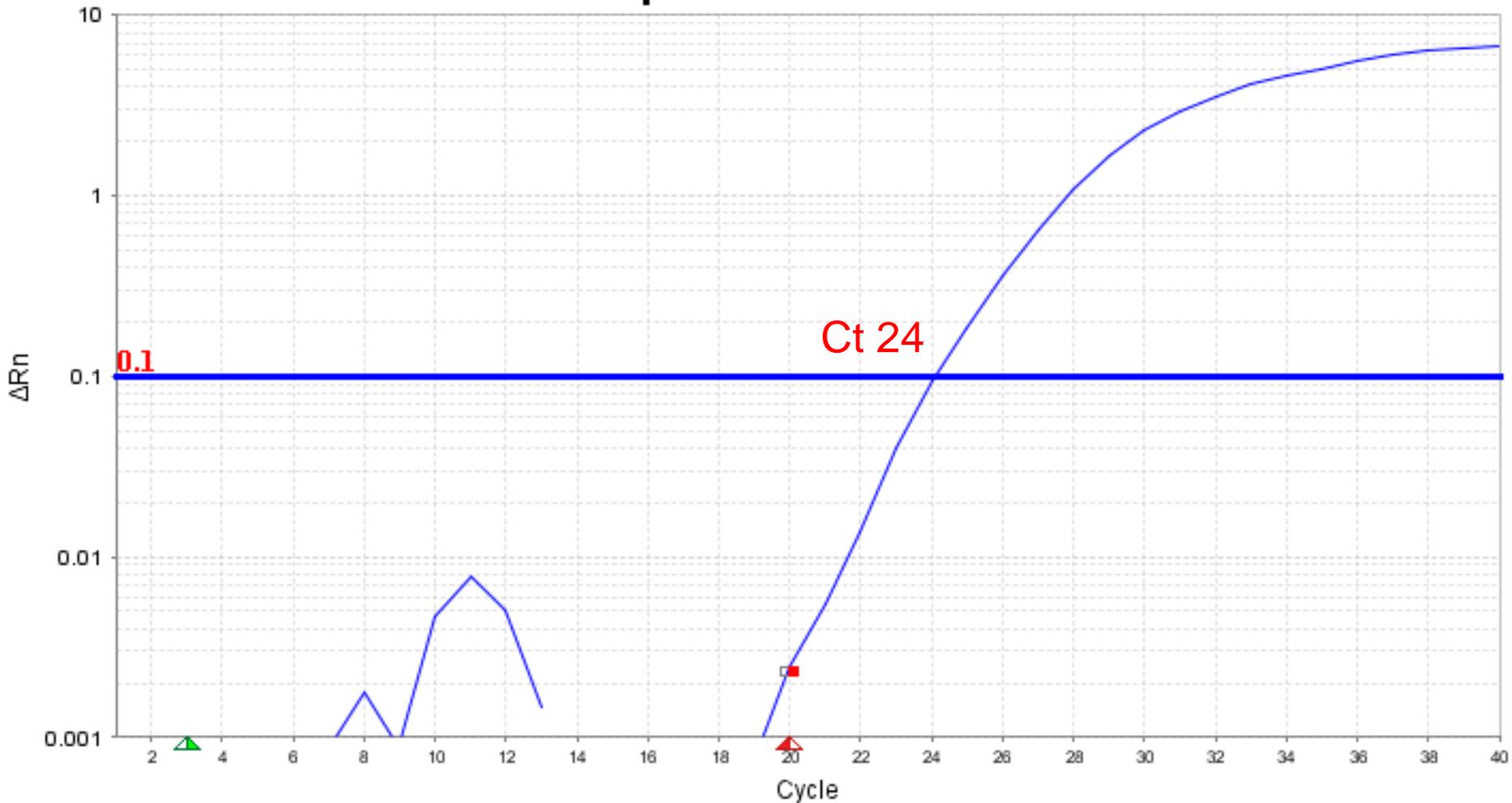
Cycle threshold (Ct)

The fractional cycle number at which each amplification curve crosses the threshold.



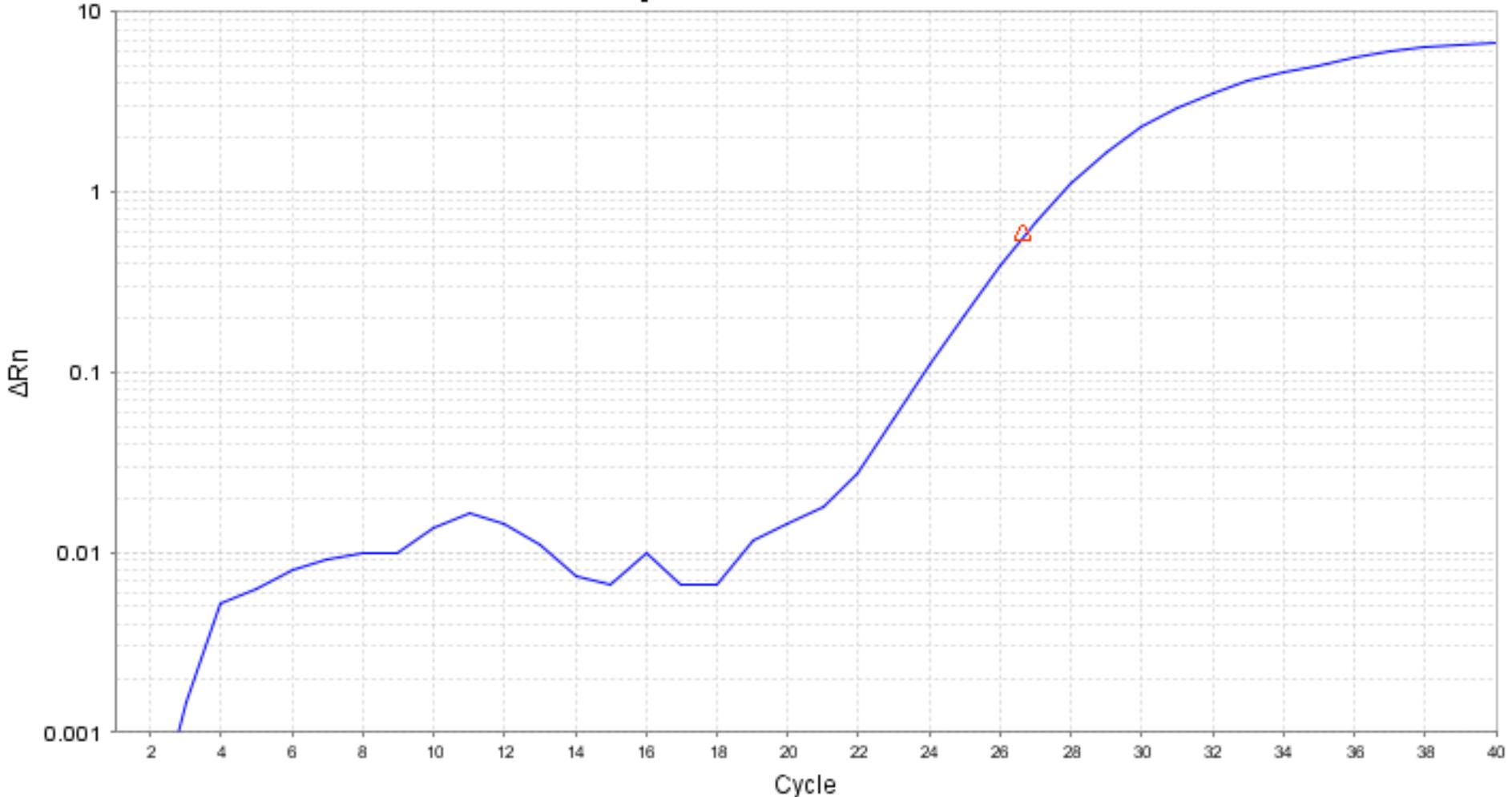
Summary of Ct algorithm (“baseline threshold”)

Amplification Plot



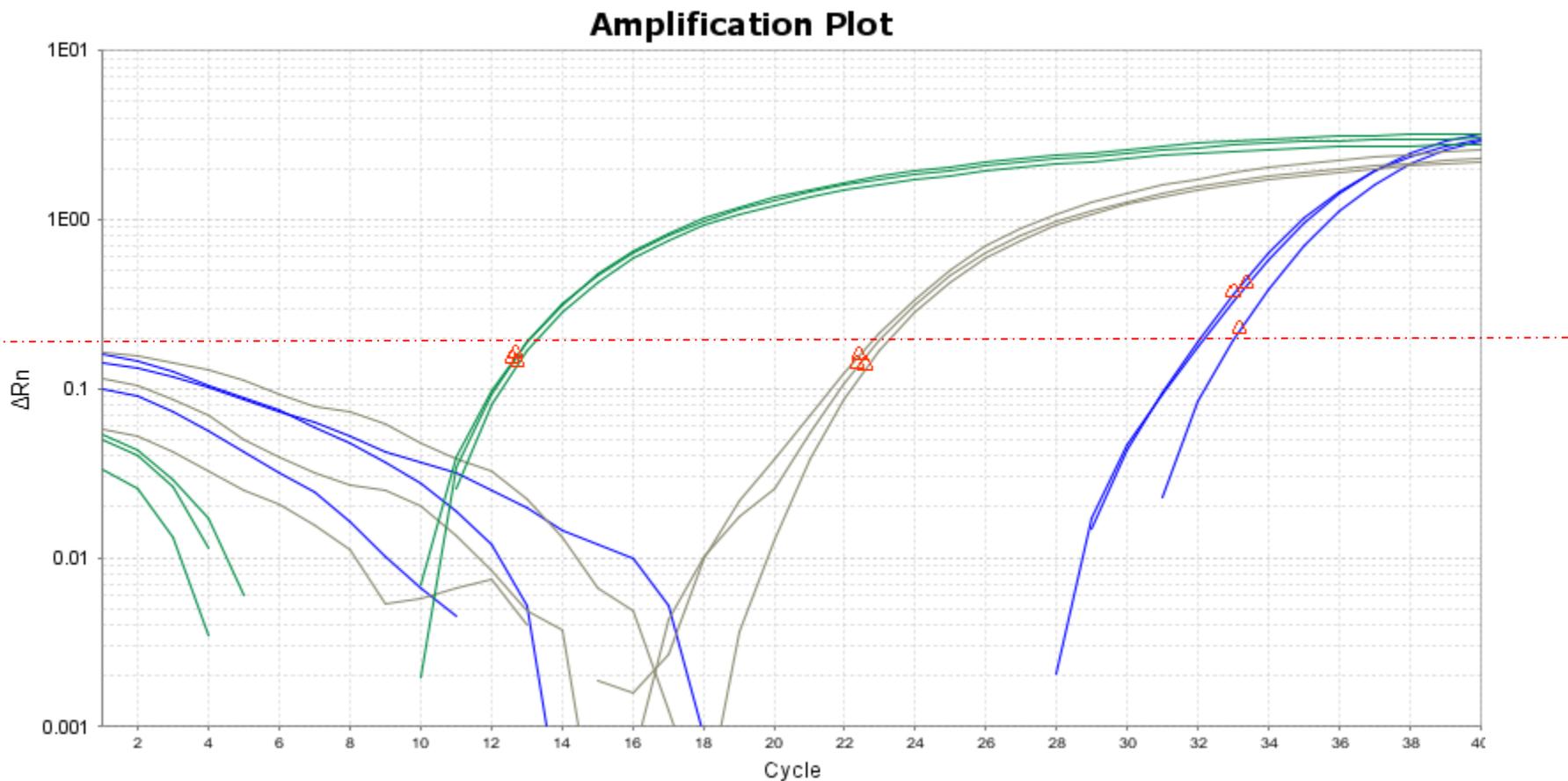
CRT algorithm (“relative threshold”)

Amplification Plot



App note: http://tools.thermofisher.com/content/sfs/brochures/CO28730-Crt-Tech-note_FLR.pdf

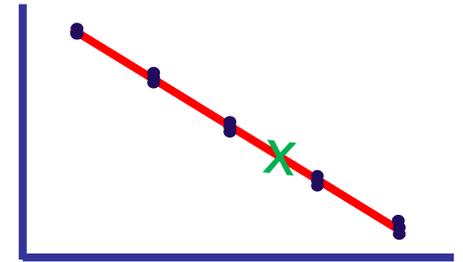
CRT may help out for samples that may need to be omitted with CT, to tighten up technical replicates.



Use Ct values to provide quantitative results

Absolute Quantitation calculates a copy number by using standard curve analysis

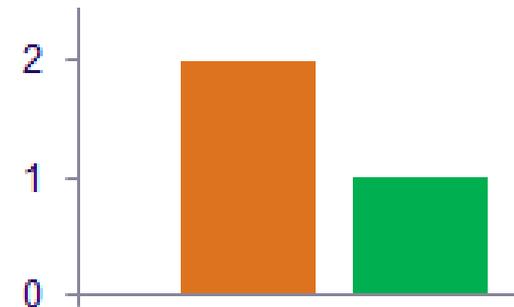
“I have 500 copies of TNF α in sample 1”



$\Delta\Delta$ Ct method calculates relative fold change

- no standard curves, assays have similar efficiencies

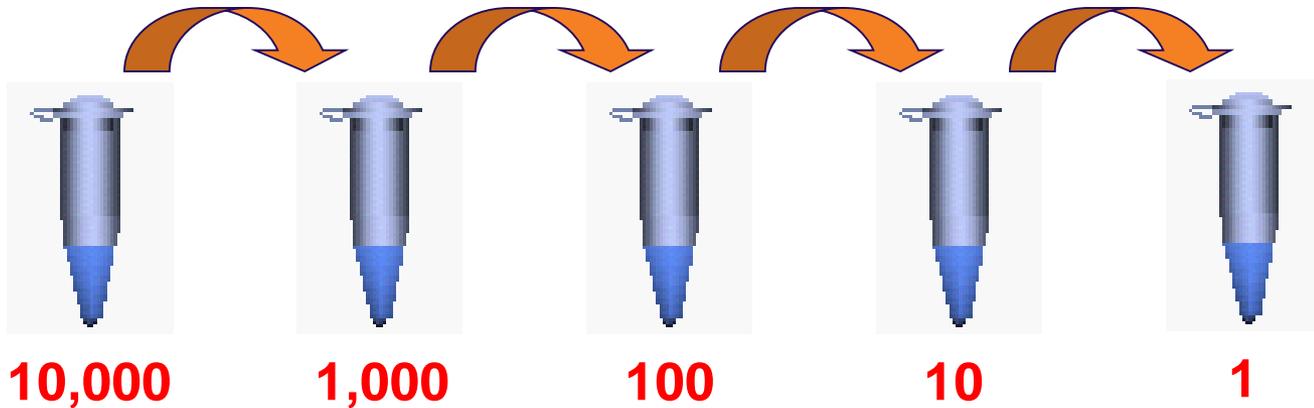
“TNF α is upregulated 2 fold in sample 1 compared to the control sample”



Generate an **absolute** standard curve

Recommend 5 or more, point standard curve of 10 fold dilutions

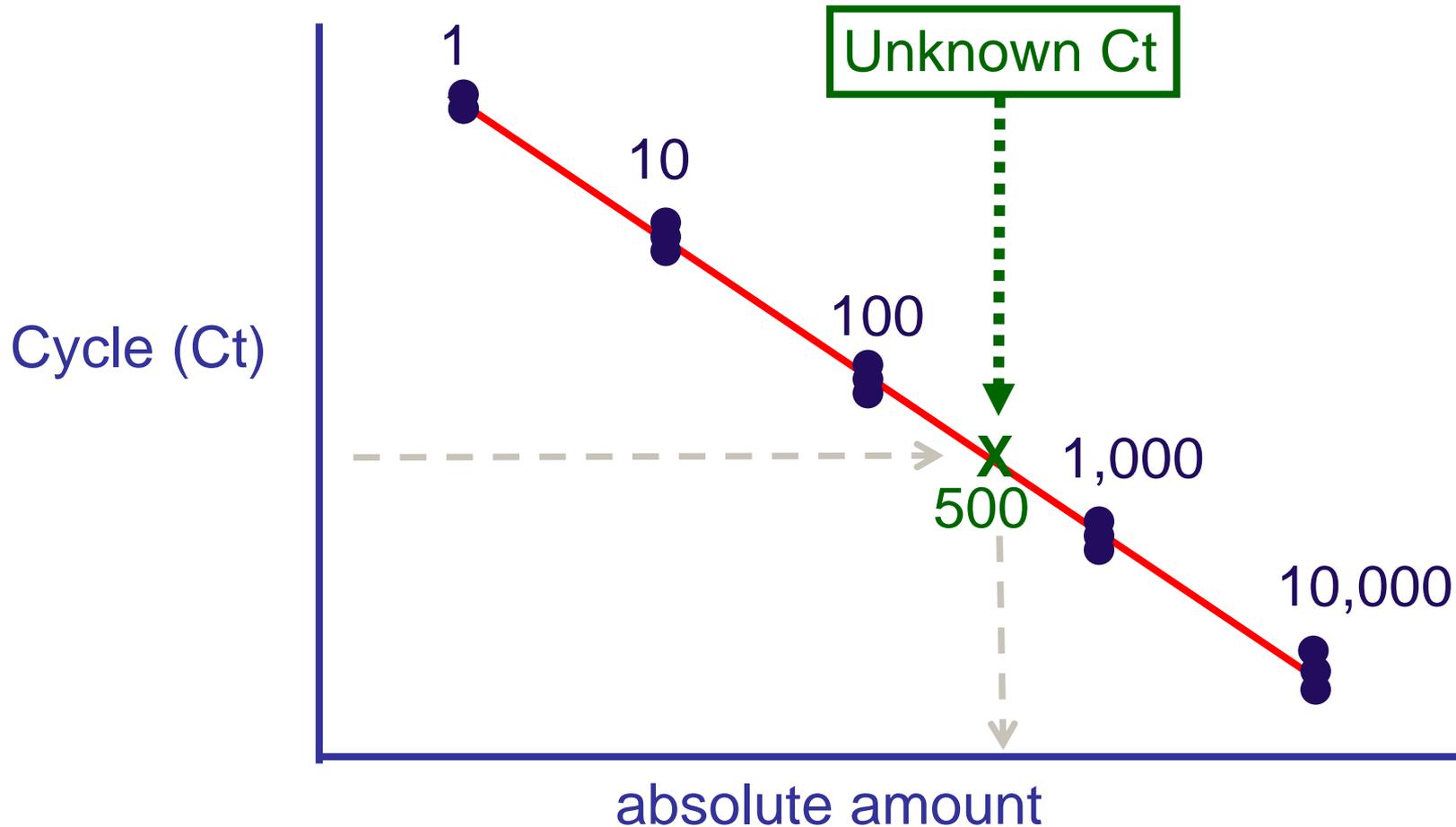
Do serial dilutions



	1	2	3	4	5	6	7	8	9	10	11	12
A	S	S	S	S	S	S	S	S	S	S	S	S
	40	40	40	30	30	30	31	34	34	37	37	37
B	U	U	U	U	U	U	U	U	U	U	U	U
	40	40	40	30	30	30	31	31	31	29	29	29

Assay

After a run, plot unknown Ct on curve



Another way to analyze expression data for relative comparisons (2^n)

ΔCt of 1 = 2^1 = 2 fold difference

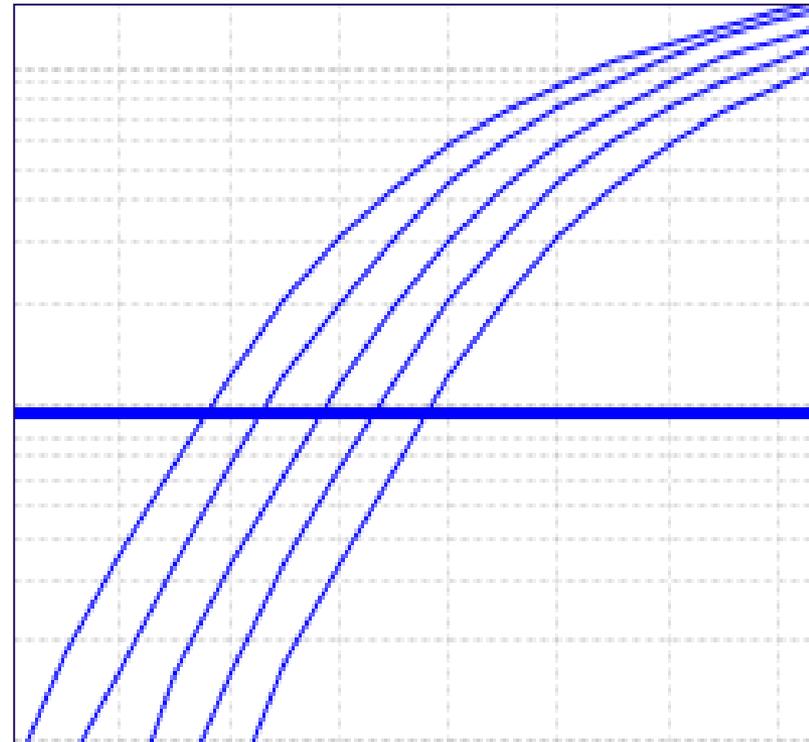
ΔCt of 2 = 2^2 = 4 fold

ΔCt of 3 = 2^3 = 8 fold

ΔCt of 3.3 = $2^{3.3}$ = 10 fold

ΔCt of 4 = 2^4 = 16 fold

This assumes that the amount of product is doubling with each cycle in the exponential phase, when it does, we call this 100% amplification efficiency.



Standard curve statistics example:

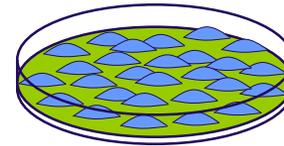
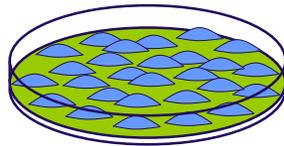
Slope: -3.384 **R²:** 0.995 **Eff%:** 97.477

$\Delta\Delta Ct$

Question: What happens to the expression of a gene when I apply a treatment?

Untreated Cells

Treated Cells



DNA/RNA Selection Guide

RNA

RNA

Dnase treatment 18068-015

SuperScript® VILO cDNA Synthesis Kit

cDNA

cDNA

gene expressed at level X

gene expressed at level Y

1 vs. 2 copies

100 vs. 200 copies \rightarrow 2 fold

Answer: Expressed as a relative fold change

In relative gene expression experiments, we examine two types of genes. . .

Gene of interest (target)

Normalizing gene (control)

A gene that is **expressed consistently** in all samples in an experiment. (a.k.a. “control gene,” “housekeeping gene,” “endogenous control”, “reference gene”)

TaqMan® Endogenous Control Array

-Human & Rodent available

-Includes 32 commonly studied control genes

-Genes range in expression from:

high, medium, low (ex. 18s, GAPDH, HPRT1)

Gene Symbols	1	2	3	4	5	6	7	8	9	10	11	12
A	18S	GAPDH	HPRT1	GUSB	18S	GAPDH	HPRT1	GUSB	18S	GAPDH	HPRT1	GUSB
B	ACTB	B2M	HMBS	IPC8	ACTB	B2M	HMBS	IPC8	ACTB	B2M	HMBS	IPC8
C	PGK1	RPLP0	TBP	TFRC	PGK1	RPLP0	TBP	TFRC	PGK1	RPLP0	TBP	TFRC
D	UBC	YWHAZ	PPIA	POLR2A	UBC	YWHAZ	PPIA	POLR2A	UBC	YWHAZ	PPIA	POLR2A
E	CASC3	CDKN1A	CDKN1B	GADD45A	CASC3	CDKN1A	CDKN1B	GADD45A	CASC3	CDKN1A	CDKN1B	GADD45A
F	PUM1	PSMCA4	EIF2B1	PES1	PUM1	PSMCA4	EIF2B1	PES1	PUM1	PSMCA4	EIF2B1	PES1
G	ABL1	ELF1	MT-ATP6	MRPL19	ABL1	ELF1	MT-ATP6	MRPL19	ABL1	ELF1	MT-ATP6	MRPL19
H	POP4	RPL37A	RPL30	RPS17	POP4	RPL37A	RPL30	RPS17	POP4	RPL37A	RPL30	RPS17

TaqMan® Endogenous Control Array

(Human & Rodent available)

	Sample 1				Sample 2				Sample 3			
Gene Symbols	1	2	3	4	5	6	7	8	9	10	11	12
A	18S	GAPDH	HPRT1	GUSB	18S	GAPDH	HPRT1	GUSB	18S	GAPDH	HPRT1	GUSB
B	ACTB	B2M	HMBS	IPC8	ACTB	B2M	HMBS	IPC8	ACTB	B2M	HMBS	IPC8
C	PGK1	RPLP0	TBP	TFRC	PGK1	RPLP0	TBP	TFRC	PGK1	RPLP0	TBP	TFRC
D	UBC	YWHAZ	PPIA	POLR2A	UBC	YWHAZ	PPIA	POLR2A	UBC	YWHAZ	PPIA	POLR2A
E	CASC3	CDKN1A	CDKN1B	GADD45A	CASC3	CDKN1A	CDKN1B	GADD45A	CASC3	CDKN1A	CDKN1B	GADD45A
F	PUM1	PSM04	EIF2B1	PES1	PUM1	PSM04	EIF2B1	PES1	PUM1	PSM04	EIF2B1	PES1
G	ABL1	ELF1	MT-ATP6	MRPL19	ABL1	ELF1	MT-ATP6	MRPL19	ABL1	ELF1	MT-ATP6	MRPL19
H	POP4	RPL37A	RPL30	RPS17	POP4	RPL37A	RPL30	RPS17	POP4	RPL37A	RPL30	RPS17



Hs99999902_m1

Example of $\Delta\Delta C_t$ Math

<u>Sample</u>	<u>X</u>	<u>N</u>
Treated 1	24	14
Treated 2	20	11
Treated 3	28	12
Untreated	24	13

X = Target N = Normalizing gene

Example of $\Delta\Delta\text{Ct}$ math

Sample	X	N	ΔCt
Treated 1	24	- 14	= 10
Treated 2	20	- 11	= 9
Treated 3	28	- 12	= 16
Untreated	24	- 13	= 11

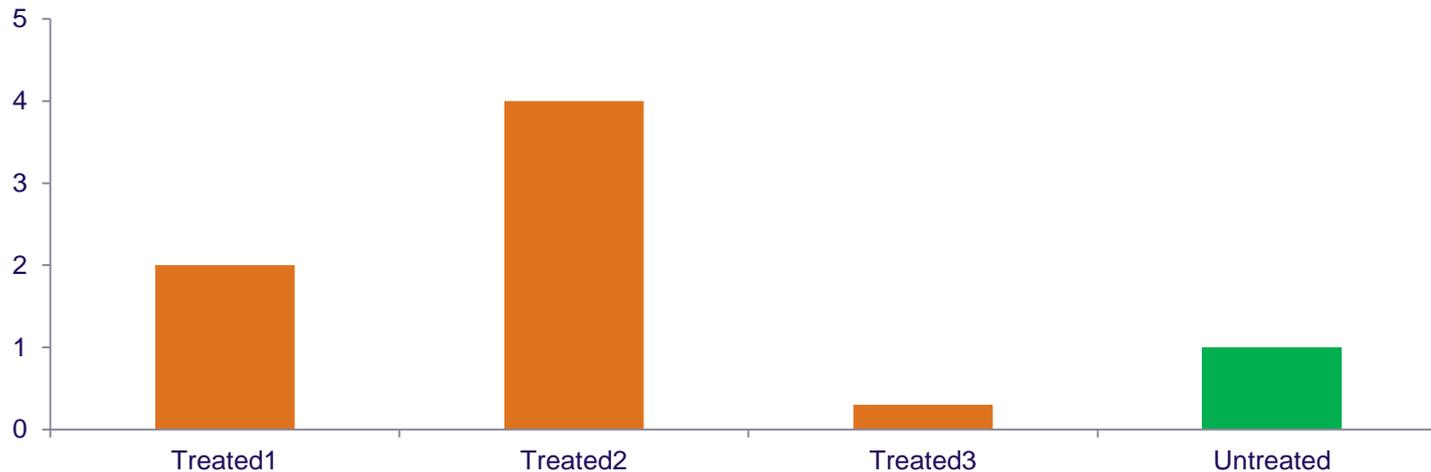
Choose a **calibrator**; all samples are relatively expressed to this sample

Sample	X	N	ΔCt	$\Delta\Delta\text{Ct}$
Treated 1	24	14	10	$-11 = -1$
Treated 2	20	11	9	$-11 = -2$
Treated 3	28	12	16	$-11 = 5$
Untreated	24	13	11	$-11 = 0$

Example of $\Delta\Delta\text{Ct}$ math

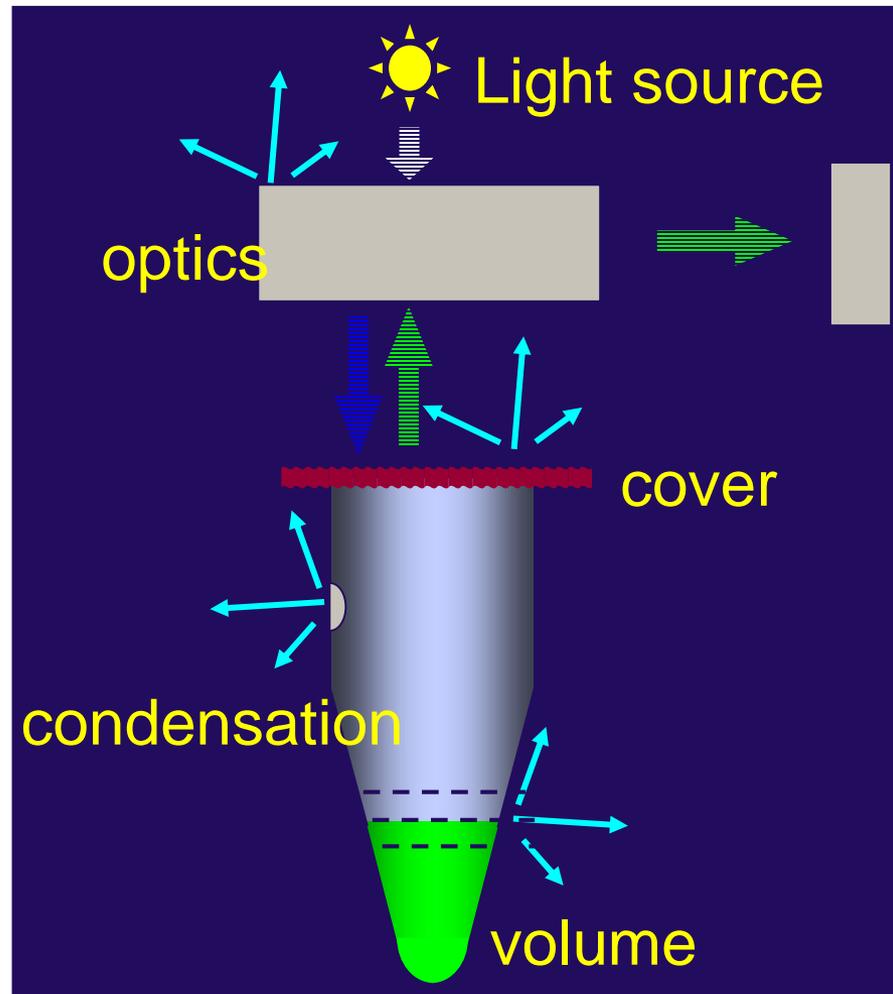
RQ (fold change)

Sample	X	N	ΔCt	$\Delta\Delta\text{Ct}$	$2^{-\Delta\Delta\text{Ct}}$	
Treated 1	24	14	10	-1	2	↑
Treated 2	20	11	9	-2	4	↑
Treated 3	28	12	16	5	0.3	↓
Untreated	24	13	11	0	1	



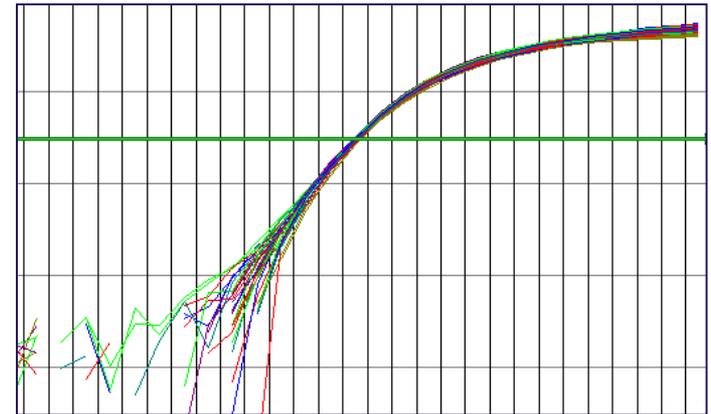
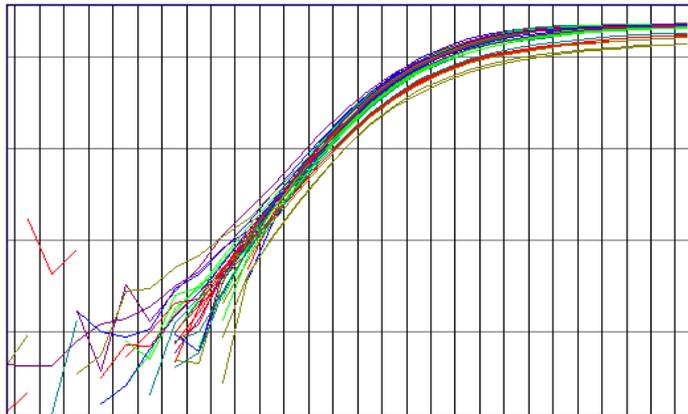
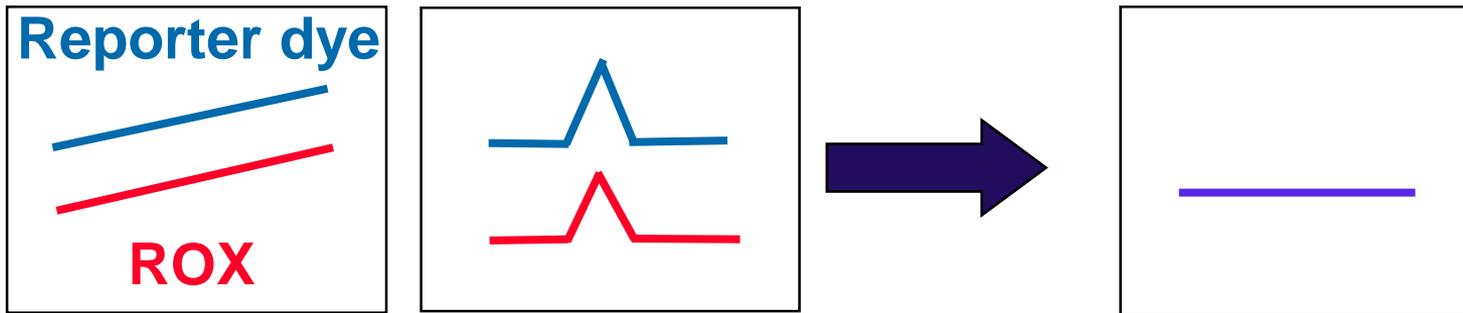
Software can perform the calculations! **ThermoFisher**
SCIENTIFIC

Other Tips & Information



ROX™ Passive Reference Dye

Improves precision of replicates by normalizing for non-PCR related variations.

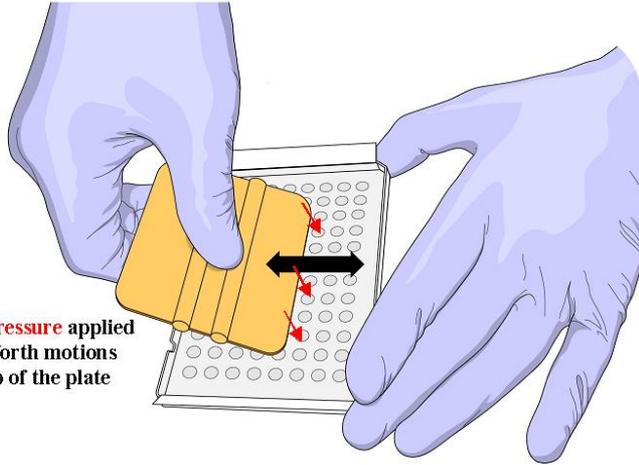


Prevent evaporation & bubbles during PCR

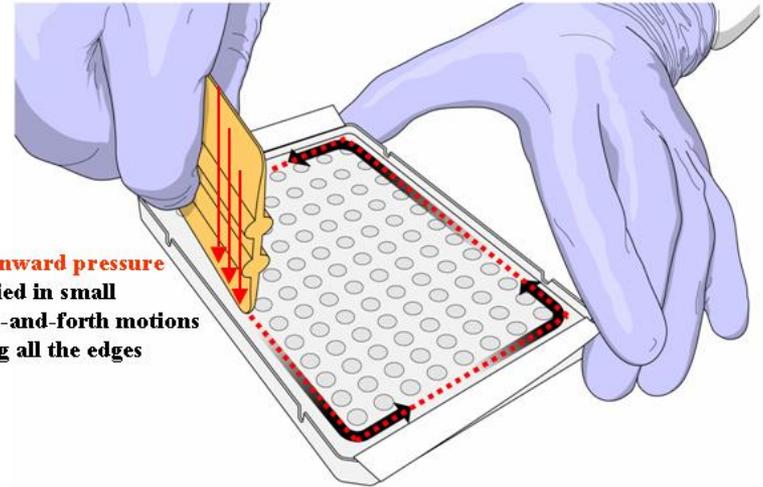
Use adhesive covers . . .

Applicator
#4333183

Downward pressure applied
in back-and-forth motions
across the top of the plate



Downward pressure
applied in small
back-and-forth motions
along all the edges



SPIN PLATES before loading into instrument!!!



Guides to multiplexing

Real-time multiplexing

Part 1 - What is multiplexing? How does it work?

Multiplexing is the amplification of more than one target in one tube or well on a plate. Duplexing is specifically the amplification of two targets in one tube, triplexing is the amplification of three targets, and so on. Multiplexing is possible with TaqMan® probe-based assays, where each assay has a specific probe, and that probe is labeled with a different colored dye. The instruments can detect the different dyes and measure the signal from each one separately, and use that information to quantitate the amounts of different targets.

How many different dyes can the instruments detect?

The number of different dyes that can be detected varies by instrument. The Applied Biosystems 7500 Real-Time PCR System can detect up to 5 different dyes, and the 7000, 7300, 7700 and 7900 can detect up to 4. However, AB prefers to reserve one of these dyes for a passive reference dye such as ROX™ dye. See Figure 1 and Figure 2.

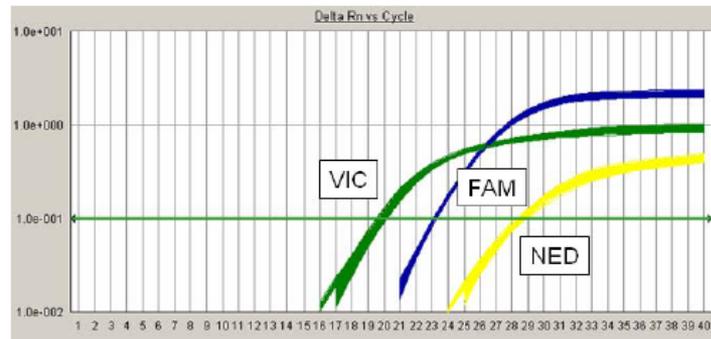


Figure 1 – Three color multiplexing on the AB 7300 instrument. Data is normalized to a fourth dye, ROX™ dye, as a passive reference.

User Bulletin #5

ABI PRISM® 7700 Sequence Detection System

August 10, 1998 (updated 01/2001)

SUBJECT: Multiplex PCR with TaqMan® VIC Probes

Overview Applied Biosystems now offers probes constructed with the new TaqMan® VIC reporter dye. The characteristics of the VIC dye make it an excellent candidate to replace existing TaqMan® JOE and HEX reporter dyes. The increased signal strength and improved spectral resolution also make VIC-labeled probes the ideal second probe for a multiplex PCR system.

This user bulletin describes the characteristics of VIC probes in relation to the existing JOE probes. It also contains guidelines for defining limiting primer concentrations in a one- or two-step multiplex reverse transcription-polymerase chain reaction (RT-PCR) system using VIC probes.

The following topics are covered in this user bulletin:

Topic	See Page
Characteristics of TaqMan VIC Probes	2
Multiplex RT-PCR	5
Technical Support	13

IMPORTANT To use VIC probes on the ABI Prism® 7700 Sequence Detection System (SDS), you must first calibrate the instrument with the Sequence Detection Systems Spectral Calibration Kit (P/N 4305822). This kit contains the new SYBR® Green and VIC fluorescent dye standards used to update the spectra components file in the SDS software. See *User Bulletin #4: Generating New Spectra Components* (P/N 4306234).

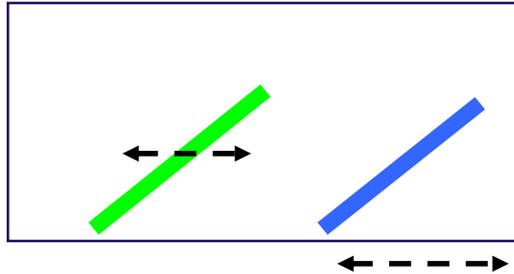
Note All documents referred to in this user bulletin are available through the Internet at the Applied Biosystems technical support documentation library or through Fax-on-Demand (see "To Obtain Documents on Demand" on page 16 for information).

The technical support documentation library is located at:

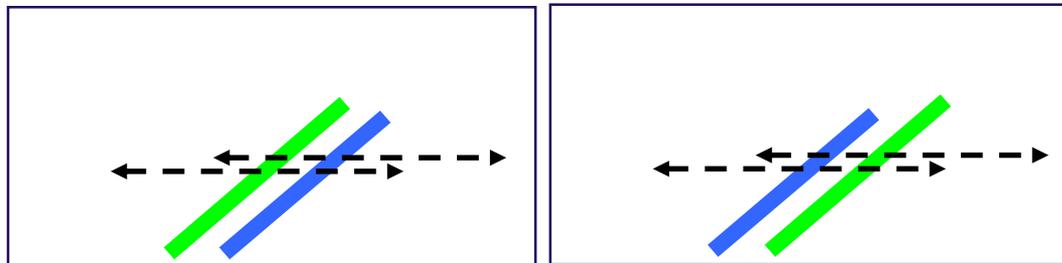
♦ www.appliedbiosystems.com/techsupport

Multiplex Scenarios

#1: One gene is more abundant.

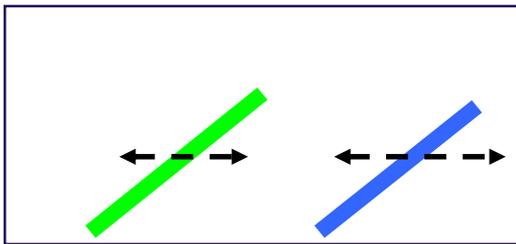


#2: Both genes are similar in expression

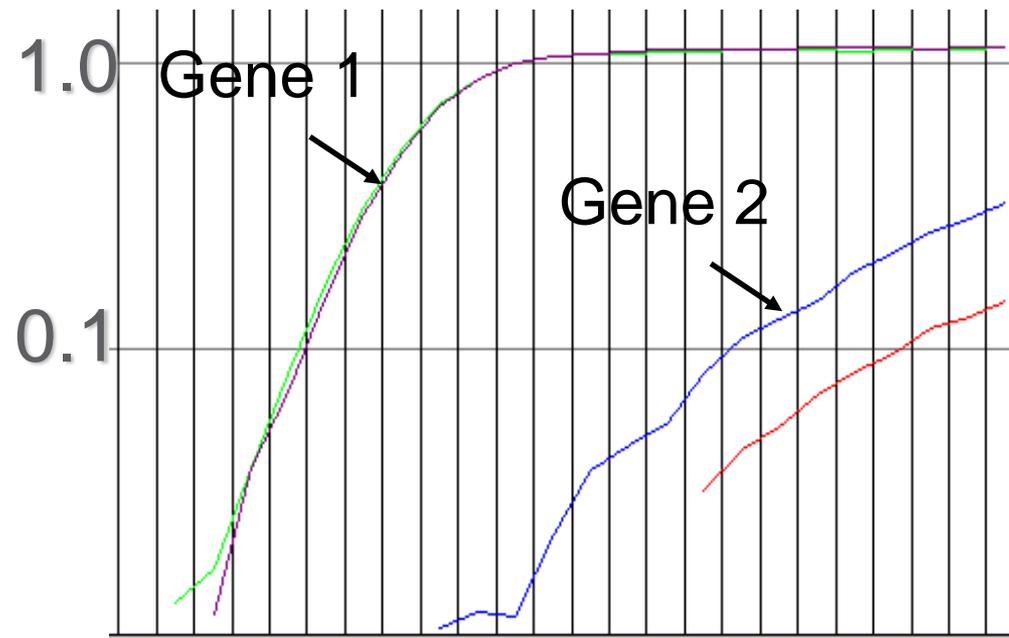


#1: One gene is more abundant

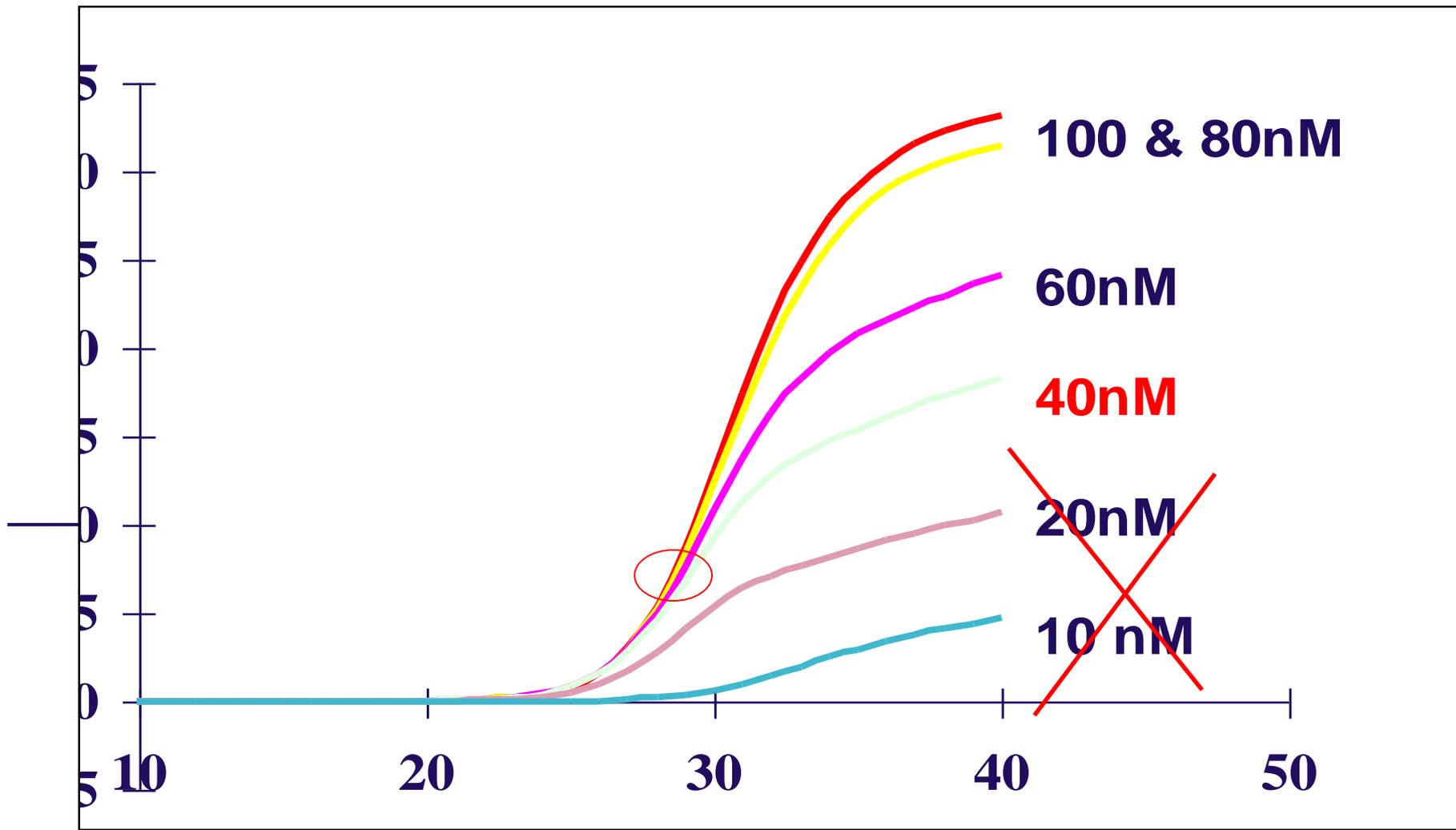
one gene uses all the reagents and there is nothing left for the other gene



high [primer] \rightarrow high plateau

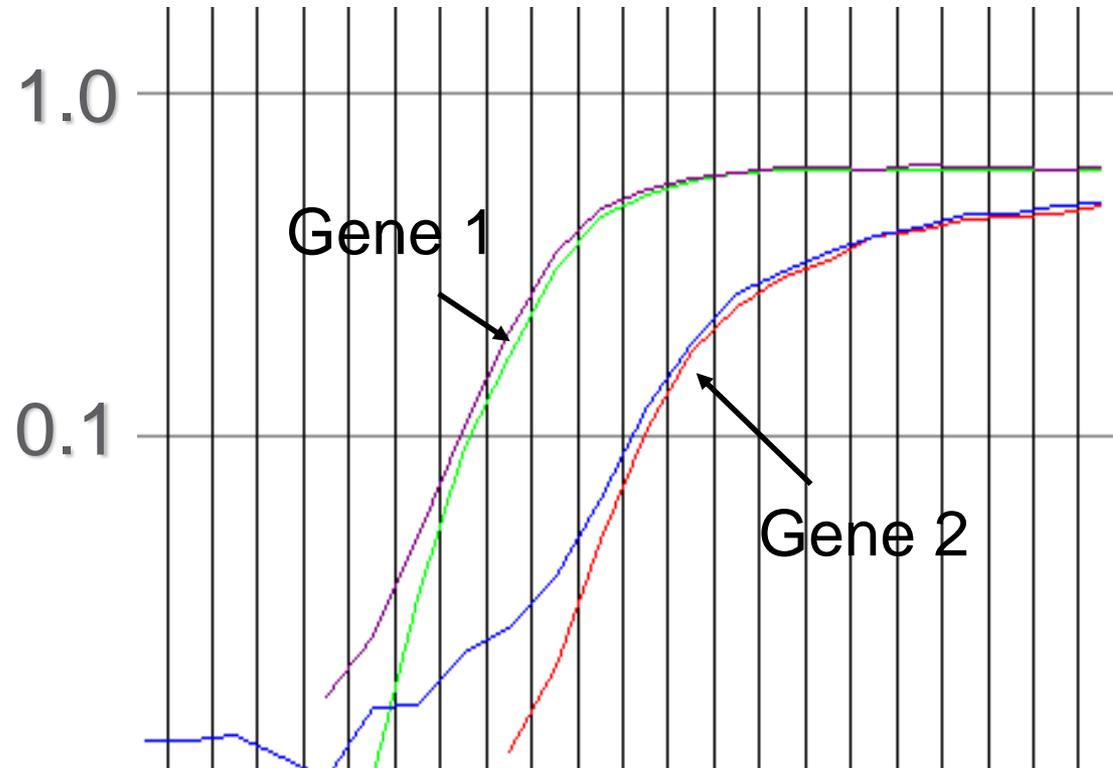


Reduce primers of more abundant gene-Optimization



Solution:

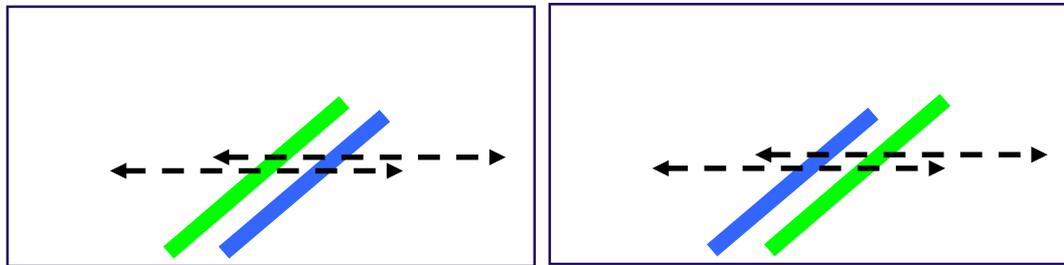
lower the [primer] of the early expressed assay →
reaction plateau quickly



- Exponential phase intact for both assays
- “primer limited” assays
- Order primers & probe separately

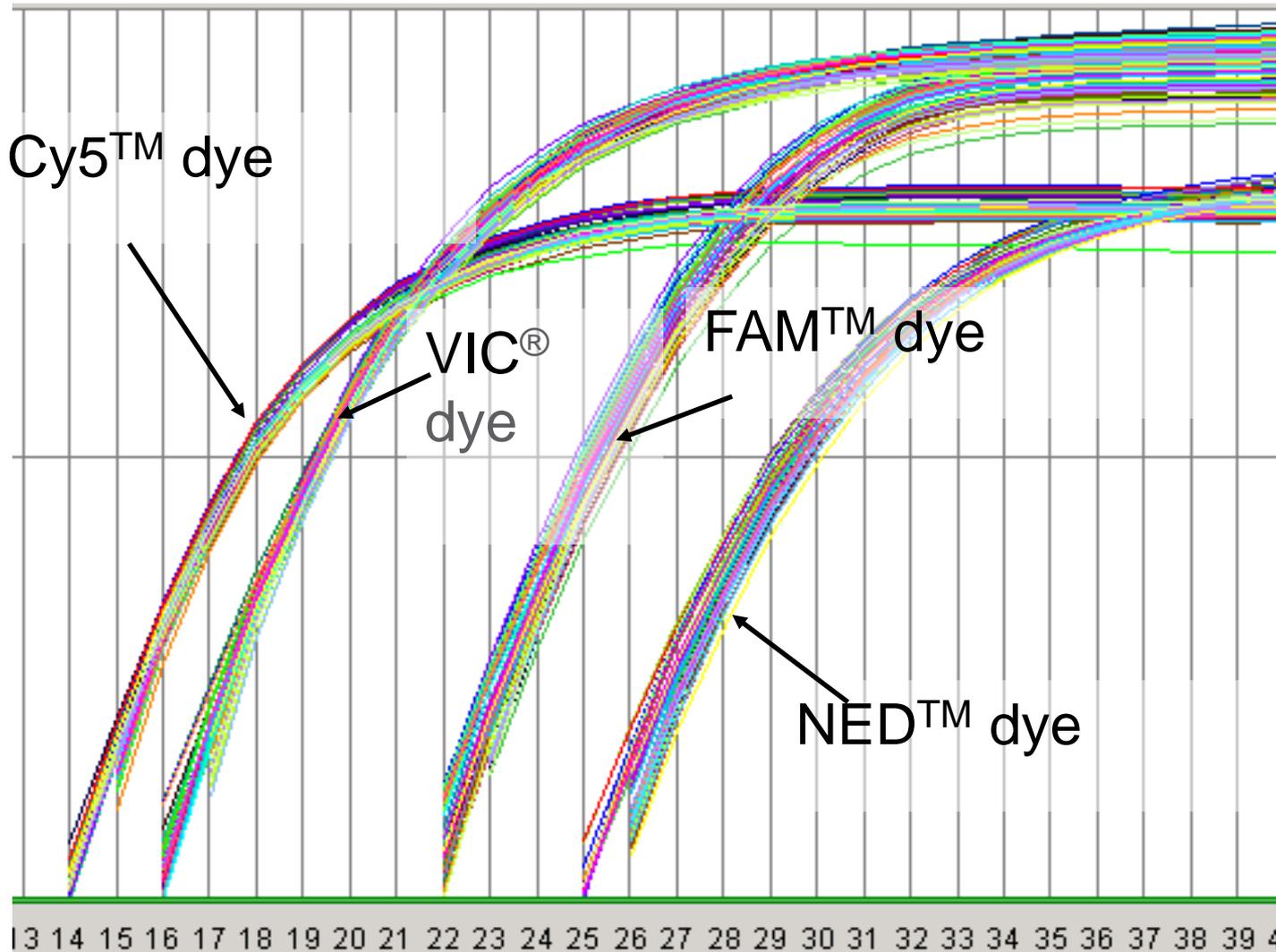
#2: Low stress multiplexing

both genes are similar in relative abundance



- Run in singleplex then duplex and ensure valid exponential phases and CTs are achieved
- If validation fails, primer limit one or both assays

Multiplexing becomes exponentially difficult with every additional assay (triplex, quadplex...)



Service Plans

Please contact Scott.Gardner@thermofisher.com for more details

Your qPCR system includes a 1 year manufacturers warranty. If a service plan is purchased, it will lock you into today's pricing and will start at the conclusion of the manufacturers warranty.

Discounts are available for multi-year coverage, locks you in at today's pricing.

Benefits of AB Assurance Service Plan

- Reduces downtime by providing proactive maintenance service
- Includes parts, labor, & travel at no additional cost
- Priority on-site guaranteed 2-day response time, & priority access to remote service engineer
- Scheduled on-site planned maintenance (PM) includes:
 - Calibration services (cost of the calibration kit is included)
 - Additional tests to ensure system performance
 - Computer repair and replacement



Support

(800) 955-6288

techsupport@thermofisher.com

Scott.gardner@thermofisher.com

Instrument software

Computer login (default),

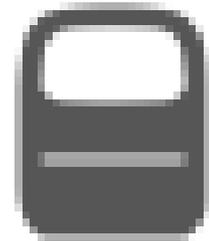
user name & password are the same

INSTR-ADMIN INSTR-ADMIN



Free Software Download (link in follow-up email)

- .edt template
- .eds single data file



Merge multiple .eds files in ThermoFisher Connect or TaqMan Genotyper (for SNP data)